Glypican-3–Specific CAR T Cells Coexpressing IL15 and IL21 Have Superior Expansion and Antitumor Activity against Hepatocellular Carcinoma

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ABSTRACT

Hepatocellular carcinoma (HCC) is the fourth most common cause of cancer-related death in the world, and curative systemic therapies are lacking. Chimeric antigen receptor (CAR)–expressing T cells induce robust antitumor responses in patients with hematologic malignancies but have limited efficacy in patients with solid tumors, including HCC. IL15 and IL21 promote T-cell expansion, survival, and function and can improve the antitumor properties of T cells. We explored whether transgenic expression of IL15 and/or IL21 enhanced glypican-3–CAR (GPC3-CAR) T cells’ antitumor properties against HCC. We previously optimized the costimulation in GPC3-CARS and selected a second-generation GPC3-CAR incorporating a 4-1BB costimulatory endodomain (GBBz) for development. Here, we generated constructs encoding IL15, IL21, or both with GBBz (15.GB dz, 21.GB dz, and 21.15.GB dz, respectively) and examined the ability of transduced T cells to kill, produce effector cytokines, and expand in an antigen-dependent manner. We performed gene-expression and phenotypic analyses of GPC3-CAR T cells and CRISPR-Cas9 knockout of the TCF7 gene. Finally, we measured GPC3-CAR T-cell antitumor activity in murine xenograft models of GPC3+ tumors. The increased proliferation of 21.15.GB dz T cells was at least in part dependent on the upregulation and maintenance of TCF-1 (encoded by TCF7) and associated with a higher percentage of stem cell memory and central memory populations after manufacturing. T cells expressing 21.15.GB dz had superior in vitro and in vivo expansion and persistence, and the most robust antitumor activity in vivo. These results provided preclinical evidence to support the clinical evaluation of 21.15.GPC3-CAR T cells in patients with HCC.

Introduction

Hepatocellular carcinoma (HCC) is the fourth most common cause of cancer-related death in the world (1). Lack of curative therapies for unresectable and/or metastatic disease, which occurs in the majority of newly diagnosed cases, results in dismal prognoses (1). Chimeric antigen receptor (CAR)–expressing T cells show clinical successes for the treatment of CD19-positive hematologic malignancies (2–6). In contrast, CAR T cells demonstrate only modest antitumor activity in patients with solid tumors, including HCC, in part due to their limited expansion and persistence (7–12). As the overall therapeutic efficacy of CAR T cells strongly correlates with their expansion and persistence in patients with CD19-positive malignancies (2, 4), translational approaches to enhance these properties may improve the antitumor efficacy of CAR T therapy in patients with HCC.

Human interleukin 15 (IL15) and IL21 are required for optimal T-cell activation, expansion, differentiation, and function (13, 14). These cytokines are absent in the HCC microenvironment, depriving T cells of survival signals (13, 14). In preclinical models of CD19+ malignancies, neuroblastoma or gliomas, CAR T cells coexpressing either IL15 or IL21 controlled tumors significantly better than CAR T cells alone (15–18). Additionally, IL15 and IL21 can synergistically promote antigen-dependent T-cell expansion and cytolytic function (19, 20). However, whether IL15, IL21, or their combined expression enhance the antitumor effector function of CAR T cells against HCC remains unknown. We previously systematically evaluated the in vitro and in vivo activity of T cells expressing CARs targeting glypican-3 (GPC3), an antigen expressed in over 70% of HCCs but not in nonmalignant tissues (21–27). We selected the GPC3-CAR containing a 4-1BB costimulatory endodomain—"GBBz”—for further clinical development, as this receptor induces favorable TH1-polarized effector cytokine release upon tumor cell engagement and produced superior expansion and antitumor activity (27). Here, to determine the impact of IL15 and IL21 on T-cell survival, persistence, and antitumor activity in preclinical models of HCC, we coexpressed IL15, IL21, or both with the GBBz GPC3-CAR in T cells.

We demonstrated that GBBz GPC3-CAR T cells coexpressing IL15 and/or IL21 specifically and effectively killed GPC3-positive tumor cells, including HCC, in an antigen-dependent manner. Our results also indicated that constitutive transgenic expression of both IL15 and IL21 together enriched for less-differentiated T cells, which were better protected from apoptosis during repeated exposures to tumor cells. We showed that combined IL15 and IL21 expression maintained the expression of T-cell factor-1 (TCF-1), a transcription factor critical for T-cell development and survival. CRISPR-Cas9–mediated knock-out of TCF7 (the gene encoding TCF-1) in IL15 and IL21 coexpressing GPC3-CAR T cells eliminated the improvement in proliferative capacity. Finally, GPC3-CAR T cells coexpressing IL15 and IL21 exhibited the most robust peak expansion and sustained persistence.
in vivo and mediate superior tumor control and survival of HCC tumor-bearing mice compared with either cytokine alone or controls. These results provided a strong rationale to evaluate IL15 and IL21 in combination in the past year. Cells were regularly tested and were negative for Mycoplasma contamination. The HCC cell line Hep3B was a kind gift from Dr. Xiao-Tong Song (Bayor College of Medicine, Houston, TX) and was tested in Dulbecco modified Eagle medium, and its identity was confirmed at the Characterized Cell Line Core Facility at the University of Texas MD Anderson Cancer Center (Houston, TX) by short tandem repeat method. Cells were obtained between 2014 and 2015. A549-GPC3 cells were generated by transducing A549 cells with a retroviral vector encoding GPC3: Huh-7 firefly luciferase (Fluc) cells were similarly generated using an Fluc/FIuc construct (27).

Materials and Methods

Cell lines
The HCC cell line Hep3B, rhabdoid tumor cell line G401, lung carcinoma cell line A549, and human embryonic kidney cell line 293T were obtained from the American Type Culture Collection and were cultured in Dulbecco’s modified Eagle media with 10% fetal bovine serum (FBS) except for 293T cells, which were cultured in Iscove modified Dulbecco media with 10% FBS. Cells were kept in culture for less than 3 weeks for any given experiment. Cells were not reauthenticated after 3 months of reimplantation with frozen stocks.

Generation of retroviral constructs
The codon-optimized minigene encoding cytokines IL21, IL15, and GPC3-CAR (“GBBz” (27) linked with T2A sequence and flanked by Ncol and Mlu restriction sites was synthesized using the GeneArt system (Thermo Fisher Scientific). This fragment was subcloned into the pSGF retroviral vector yielding the 21.15.GBBz retroviral construct (Fig. 1A). 21.GBBz was generated from 21.15.GBBz by PCR amplifying genes encoding IL21 (F: ATCCTCTAGACTGCATGGAACGGATC/R: CAGTGCGGCCGCTCAGGCCCTGCTGGTGTT) and in subsequent In-Fusion cloning (Fig. 1A). The 21.15 construct was produced by PCR amplifying the fragment encoding both IL21 and IL15 from 21.15.GBBz (F: ATCCTCTAGACTGCATGGAACGGATC/R: CAGTGCGGCCGCTCAGGCCCTGCTGGTGTT) and in the process generating NcoI and NotI restriction enzyme sites in the introduced 5’ and 3’ overhangs. This fragment was cloned into a pSGF plasmid bearing the orange monomeric derivative of DsRed (28). Sequencing was performed following each cloning step and confirmed the alterations were made (Epoch Life Science).

Retrovirus production and transduction of primary T cells and cell lines
Transient transfection of HEK 293T cells with plasmids for GPC3-CAR constructs, RFP plasmid encoding the RD114 envelope, and PegPam3 plasmid encoding the MoMLV gag-pol was used to generate retroviral supernatants as previously described (30). OKT-3/CD28 mAb-coated plates were used to stimulate human peripheral blood mononuclear cells isolated from healthy volunteer donors (Gulf Coast Regional Blood Center). After 48 hours, cells were washed and replated in complete RPMI medium and maintained in incubators at 37 °C with 5% CO2 (27).

Cytotoxicity assay
Cytotoxicity of GPC3-CAR T cells was assessed as described previously (27) using a standard 4-hour chromium 51 (Cr) release assay. Briefly, target cells were labeled with 51Cr for 1 hour followed by incubation with effector cells for 4 hours at 37°C using multiple effector-to-target (E:T) ratios. Cell culture supernatants were collected, and radioactivity was measured in a gamma counter (PerkinElmer).

Measurement of cytokines and chemokines
The Human IL15/IL21 ELISA MAX Deluxe kit (BioLegend) was used to measure IL15 and IL21 concentrations according to the manufacturer’s instructions. Briefly, 0.5 × 106 resting CAR T cells were cultured in the presence or absence of Huh-7 cells at a 1:1 ratio. Cell culture supernatants were collected at 72 hours, centrifuged, and frozen until the time of assay. Multiplex cytokine/chemokine immunoassays were performed as previously described using the MILLI-PLEX MAP human cytokine/chemokine magnetic bead kit (EMD Millipore, cat. #HCYTOMAG-60K) according to the manufacturer’s instructions (27).

To assess the production of IL1b and IL6, monocytes negatively selected from human PBMCs by Pan monocyte isolation kit (Miltenyi Bio 130-096-537 kit) were plated at 2.5:2.5:1 tumor, CAR T-cell, and monocyte ratio, respectively. Monocyte purity was confirmed by FACS and was above 95% for each coculture experiments. Supernatants were collected at 20 hours and evaluated with IL1b and IL6 ELISA kits (BioLegend, cat. #437005 and R&D Systems, cat. #D6050, respectively) according to the manufacturer’s manual.

Flow cytometry
GPC3-CAR expression was detected using the anti-F(ab)2 Alexa Fluor 647-conjugated antibody (Jackson ImmunoResearch) and anti-goat IgG1 isotype control (Jackson ImmunoResearch). The following antibodies were used for T-cell phenotyping analyses: anti-CD4-APC/Flur 750 (BioLegend), anti-CD8-V500 (BD Biosciences), anti-CD28-PeCy7 (BioLegend), anti-CD95-FITC (BioLegend), anti-CD49d-PE/Cy7 (LAG-3; BioLegend), and anti-CD366-BV421 (TIM-3; BD Biosciences). Anti-bovine IgG (Sigma-Aldrich) was used to block nonspecific binding of other murine antibodies following CAR staining. Flow cytometry assessment was performed on either an LSR-II (BD Biosciences) or iQue Screener PLUS (Intelllicyt Corporation). Results were analyzed using FlowJo software (FlowJo). To detect intracellular TCF-1 expression, CAR T cells were first stained for surface expression of CAR, CD4, and CD8 using the antibodies listed above, followed by staining with anti-TCF1-PE (TCF1, BioLegend) used in conjunction with the True-Nuclear Transcription Factor Buffer Set (BioLegend) according to the manufacturer’s instructions.

Repeat stimulation stress test
In vitro expansion and persistence were assessed by repeatedly coculturing CAR T cells with fresh Huh-7 tumor cells every 3 to 4 days at a 1:1 ratio. At the end of each coculture interval, CAR T-cell counts were measured by flow cytometry staining for the CAR and using
Figure 1. Generation of GPC3-CAR T cells that coexpress IL21 and IL15. A, Schematic of retroviral constructs encoding GPC3-CAR (GBBz) with and without IL15 and/or IL21. B, CAR expression in T cells transduced (on day 3 after stimulation with plate-bound antibody) using retroviral vectors containing the indicated GPC3-CAR constructs as measured by flow cytometry (on days 10–14). Data from 1 representative donor and summary for 8 independent donors in independent expansions are shown (mean ± SD). C and D, IL15 and IL21 produced by the indicated T-cell groups at baseline (left) or after stimulation with GPC3-positive Huh-7 cells (right, E:T = 1:1, +72 hours) as measured by ELISA (mean ± SD, n = 8). One-way ANOVA, *P < 0.05, **P < 0.01, ***P < 0.001, ns, not significant.
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CountBright beads (ThermoFischer; cat. #C6950) according to the manufacturer's instructions. Annexin V staining was performed to assess CAR T-cell viability 2 days after each stimulation using the ApoScreen Annexin V Apoptosis Kit (SouthernBiotech; cat. #10010-09) according to the manufacturer's instructions.

Gene-expression profiling

CAR T cells were sorted using a Sony SH800Z instrument (Sony Biotechnology) and expanded for 1 week in complete RPMI supplemented with 100 units/mL penicillin, 100 μg/mL streptomycin, and 0.25 μg/mL amphotericin B (Thermo Fisher Scientific). Sorted CAR T cells were cocultured with tumor cells at a 2:1 (E:T) ratio for 3 days. To confirm the absence of tumor cells prior to RNA extraction, cocultured cells were analyzed by flow cytometry and the absence of residual tumor cells was confirmed. RNA was extracted using the RNeasy Mini Kit (Qiagen; cat. #74104) for subsequent experiments. sgRNA design and subsequent experiments. sgRNA design and TCF7 gene disruption in primary activated T cells were performed as previously described (31).

CRISPR-Cas9-mediated TCF7 knockout

We screened 6 TCF7-specific sgRNAs and based on their efficiency to knock out TCF7 (resulting in diminished expression of TCF1), the protein encoded by the gene) measured by FACS selected 2 sgRNAs targeting the TCF7 gene locus (target sequences GGGGTCCACT-GCTGAAATT and TCF7 GGGTCCACT-GCTGAAATT for subsequent experiments. sgRNA design and TCF7 gene disruption in primary activated T cells were performed as previously described (31).

In vivo experiments

All mice used in this study were maintained at the Small Animal Core Facility of Texas Children’s Hospital and handled under protocols approved by the BCM Institutional Biosafety Committee and Institutional Animal Care and Use Committee. Tumor cells or CAR T cells were diluted in 100 μL normal saline and were injected via indicated routes.

In vivo antitumor activity of infused CAR T cells and mouse survival were evaluated in murine HCC xenograft models as described previously (32) with modifications. Briefly, 12-week-old female NOD.CR-Pkdcre1ld2gpmf1Wts1/Sld (NSG, The Jackson Laboratory) mice were injected intraperitoneally (i.p.) with 2 × 106 Fluc+ Huh-7 cells or 5 × 106 Fluc− G401 cells followed by 0.5 × 106 or 2 × 106 CAR T cells in 100 μL normal saline intravenously (i.v.) via tail vein 1 or 2 weeks later, as indicated. Mice were assessed daily, and tumor bioluminescence was measured after i.v. injection of luciferin using the IVIS Lumina III imaging system (PerkinElmer).

To evaluate CAR T cell in vivo proliferation and persistence, mice were injected with 2 × 106 Huh-7 cells i.p. followed 2 weeks later by i.v. injection of 2 × 106 CAR T cells coexpressing an optimized Fliuc (32). Mice were imaged every other day following CAR T-cell injection to monitor expansion. Blood and spleens were collected on days 15 and 18, respectively, and evaluated for the presence of CAR T cells by flow cytometry. All animals were sacrificed according to institution standards prior to harvesting spleens. Cells were stained for mouse CD45 using anti-mouse CD45-PE or PerCP/Cy5.5 (both BioLegend) and human CD4, human CD8, and the GPC3-CAR as described above. IL15 and IL21 in blood plasma were measured using the MILLIPLEX MAP human cytokine/chemokine magnetic bead kit (EMD Millipore).

To measure the ability of inducible caspase-9 (iC9) to eliminate IL15- and IL21-producing cells, the construct iC9.21.15.NFGR was generated by infusion cloning. In brief, the fragment of IL21 and IL15 was amplified from 21.15.GBBz and fused in frame with the ampiclons of iC9 (33) and the truncated Neural Growth Factor Receptor (NFGR; ref. 34). T cells were cotransduced with GBBz, iC9.21.15.NFGR, and Fluc to track their in vivo expansion and persistence with bioluminescence imaging. The chemical inducer of dimerization AP20187 (Clontech Laboratories; cat. #635069) was dissolved according to the manufacturer's instructions given at 50 μg/mouse dose i.p. in 100 μL solution on days 8 and 10 after adoptive transfer.

Statistical analyses

Data were summarized using descriptive statistics. One-way or two-way ANOVAs adjusted for donor effect followed if applicable by pairwise comparisons between groups were carried out. Response variables were log-transformed, if necessary, to achieve normality. Analysis was performed using SAS version 9.4. P values < 0.05 were considered statistically significant.

Results

T cells effectively coexpressed IL21 and/or IL15 with a GPC3-CAR from a single retroviral construct

We generated a set of CAR constructs based on our previously optimized GBBz GPC3-CAR (27) with additional sequence(s) for human IL21 and/or IL15 (Fig. 1A) using the clinically validated Moloney murine leukemia virus-derived SFG retroviral vector backbone. After transduction, all constructs were stably expressed by human peripheral blood T cells, with constructs containing IL21 (21.GBBz and 21.15.GBBz) demonstrating slightly lower overall transduction efficiency compared with the GBBz construct (P < 0.001; Fig. 1B).

To measure IL15 and IL21 production, supernatants were collected from GPC3-CAR T cells cultured with and without GPC3-positive tumor cells and evaluated by ELISA. IL15 and IL21 were indeed secreted by CAR T cells engineered to express the corresponding genes at baseline (Fig. 1C and D). Following CAR stimulation by GPC3-positive HCC cells, IL15 and IL21 production increased significantly from T cells coexpressing the corresponding transgenes (P < 0.001), and IL21 concentrations remained significantly higher than IL15 (P < 0.001). Whereas T cells expressing 15.GBBz produced significantly more IL15 than 21.15.GBBz T cells at baseline (P < 0.01) and after stimulation (P < 0.001), IL21 production did not differ between 21.GBBz and 21.15.GBBz groups in either condition. Given that IL15 and/or IL21 could potentially induce antigen-independent T-cell proliferation, we evaluated the ability of each T-cell group to maintain autonomous growth. Whereas all groups underwent an initial burst of proliferation after transduction, in the absence of antigen stimulation, no viable CAR T cells remained in any group after 50 days (Supplementary Fig. S1).

In summary, transduced T cells stably expressed GPC3-CAR constructs and produced significant quantities of one or both cytokines, as appropriate, without evidence of antigen-independent autonomous growth.

IL15 and/or IL21 coexpression altered the effector cytokine production profile of GPC3-CAR T cells

We next explored whether IL21 and/or IL15 coexpression affects the efficacy and/or specificity of GPC3-CAR–mediated tumor cell killing using a chromium-51 release assay (27). T cells expressing any of the
four GPC3-CAR constructs specifically and effectively lysed GPC3-positive tumor cells (HuH-7, Hep3B, G401, A549-GPC3) in an antigen-dependent manner regardless of IL21/IL15 coexpression (Fig. 2A).

To evaluate an additional measure of T-cell activation, we determined the T_{reg} and T_{eff} effector cytokine production profiles of GPC3-CAR T cells following coculture with GPC3-positive or -negative target cells. GPC3-negative A549 cells did not induce significant effector cytokine production, suggesting that the GPC3-CARs in this study do not trigger consequential tonic signaling (Supplementary Fig. S2). CAR engagement by GPC3-positive Huh-7 cells specifically induced cytokine production by GPC3-CAR T cells but not by control groups (Fig. 2B and Supplementary Fig. S2). IL15 coexpression caused a significant decrease in IL13 production compared with T cells expressing constructs without IL15 (P < 0.001; Fig. 2B). A similar decline was observed for GM-CSF in 15.GBBz but not in 21.15.GBBz T cells (P = 0.0052). IL21 coexpression significantly enhanced IL2 production compared with groups lacking IL21 (21.15.GBBz vs. GBBz, P = 0.0047; 21.GBBz vs. GBBz, P = 0.0016).

Cytokine release syndrome (CRS) and neurotoxicity are serious and potentially lethal side effects of CAR T-cell therapies and are primarily mediated by IL1 and IL6 produced by monocyte/macrophages (35). These myeloid cells can be influenced by TNFα, INFγ, and GM-CSF, which are predictors of severe CRS (36). In coculture systems, we detected no difference in IL1 and IL6 produced by monocyte/macrophages between GPC3-CAR T-cell groups, suggesting that the coexpression of IL15 or IL21 was unlikely to increase the risk of CRS or neurotoxicity (Supplementary Fig. S3).

CD4+ and CD8+ T cells can produce different effector cytokines. CD8+ T cells have been shown to produce IFNγ and TNFα, which contribute to antitumor activity. CD4+ T cells produce IL2, IL4, IL5, and IL10, which are involved in the regulation of immune responses. The CD8+ T cells in CAR T-cell therapy are generally considered to be central memory (Tcm), which is associated with a high level of persistence and effector function. CD4+ T cells are mainly CD45RO+ (effector memory) and CD45RA+ (naïve) cells. GPC3-negative A549 cells did not induce significant changes in cytokine production by GPC3-CAR T cells. CD8+ 21.15.GBBz T cells had a significantly higher proportion of CD4+ central memory cells (T_{mem}, CD45RO+ /CD62L+; P = 0.0035 and P < 0.001, respectively) and of CD8+ stem cell memory/naïve cells (T_{stem}, CD45RO+ /CD62L−; P = 0.014 and P = 0.012, respectively; Fig. 3B). After stimulation with tumor cells, these differences were no longer detected (Supplementary Fig. S4B and S4C).

Next, we examined whether the observed differences in proliferation of CAR T cells were related to differences in rates of apoptosis using annexin V staining. We found that coexpression of IL15 alone or in combination with IL21 decreased the rate of apoptosis in T cells resulting in more live cells compared with GBBz alone following three rounds of stimulation with GPC3-positive tumor cells (P = 0.0012) at baseline and significantly higher expression of LAG-3, TIM-3, and PD-1 (P < 0.001, P = 0.001, and P = 0.018, respectively) after stimulation with HCC compared with GBBz cells, indicating an exhausted phenotype (Supplementary Fig. S5).

IL21 and IL15 coexpression maintained TCF-1 expression in GPC3-CAR T cells

To explore the mechanisms driving differences in proliferation between GPC3-CAR T-cell groups, we examined gene-expression profiles before and after exposure to GPC3-positive HCC cells. We detected significant differences in overall expression patterns at baseline (after manufacturing) in 21.GBBz and 21.15.GBBz T cells compared with GBBz T cells (day 0; Supplementary Figs. S6A–S6C and Fig. S7). Following stimulation with HCC cells (day 3), the gene-expression profiles of 15.GBBz, 21.GBBz, and 21.15.GBBz T cells were different from that of GBBz T cells (Fig. 4A and Supplementary Fig. S6D–S6F) with global differences included genes related to cytotoxicity (GZMA, GZMB, PRF1), chemotaxis (CCR1, CCR2, CCR5), and apoptosis/survival (BCL-2, TCF7, TCF1). Because IL15 and IL21 coexpressing CAR T cells had improved proliferation/survival, we focused on genes related to apoptosis and proliferation and found that BCL-2 and TCF7 were overexpressed in 15.GBBz and 21.15.GBBz compared with GBBz T cells (P = 0.023 and 0.0025, respectively; Supplementary Fig. S6D–S6F). No difference was detected at the protein level for BCL-2 (Supplementary Fig. S8), suggesting that it does not play a key role in the enhanced survival of 21.15.GBBz T cells. The TCF1 protein, encoded by TCF7, is a critical transcription factor for T-cell development, expansion, and survival (38, 39). Prior to stimulation (day 0), the TCF7 gene was expressed similarly in all the GPC3-CAR T-cell groups (Supplementary Fig. S6A–S6C). After stimulation with HCC cells, the gene expression of TCF7 was significantly increased in 21.GBBz (2.2-fold increase, P = 0.014), 15.GBBz (2.3-fold increase, P = 0.028), and to an even greater extent in 21.15.GBBz T cells (4.1-fold increase, P = 0.0003) compared with GBBz T cells (Fig. 4A). The proportion of TCF1 (protein)–positive cells at baseline was significantly higher in 21.15.GBBz T cells compared with other CAR T-cell groups (21.15.GBBz vs. 15.GBBz in CD4 subset: P = 0.0223; in CD8 subset: P < 0.001; Fig. 4B and C). The expression of either IL15, IL21, or the...
Coexpression of IL15 and/or IL21 maintained GPC3-specific tumor cell killing but altered effector cytokine release in GPC3-CAR T cells. A, GPC3-CAR T cells were cocultured with ⁵¹Cr-labeled GPC3-positive (Huh-7, Hep3B, G401, and A549-GPC3) and -negative (A549) target cells at the indicated E:T ratios. ⁵¹Cr-release was detected after 4 hours as a measure of GPC3-CAR T cytotoxicity (mean ± SEM, pooled results from two independent experiments evaluating 4 donors). B, Indicated T-cell groups were cultured with Huh-7 tumor cells at a 1:1 ratio, and concentration of indicated effector cytokines released into the supernatant (+ 24 hours) was measured by Luminex assay (combined data from three independent experiments evaluating 6 donors). Comparison with two-way ANOVA. C, Surface expression of CD4 (top) and CD8 (bottom) populations within CAR-positive cells at baseline (left) and after two consecutive stimulations with Huh-7 cells (right, E:T = 1:1) as measured by FACS (mean ± SD, combined data from three independent experiments evaluating 6 donors), one-way ANOVA. *, P < 0.05; **, P < 0.01; ***, P < 0.001.
Coexpressing IL21 and IL15 enhanced expansion, enriched for less-differentiated subsets, and reduced the apoptosis rate of GPC3-CAR T cells. 

**A,** GPC3-CAR T cells were cocultured with HCC cells at a 1:1 ratio and replated every 3 to 4 days as indicated with fresh HCC cells in the absence of exogenous cytokines (mean ± SEM, combined data from three independent experiments evaluating 3 donors). One-way ANOVA followed.

**B,** Phenotype of GPC3-CAR T cells as measured by surface expression of CD45RO and CD62L after manufacture. Shown are representative flow plots and summary data for indicated CAR T-cell groups (mean ± SEM, combined data from four independent experiments evaluating 4 donors; asterisks indicate significant differences from GBBz). Tscm/Tn: CD45RO⁻/CD62L⁺, Tcm: CD45RO⁺/CD62L⁺, Tem: CD45RO⁺/CD62L⁻, T eff: CD45RO⁻/CD62L⁻. Tscm, effector memory T cell; Teff, effector T cell.

**C,** GPC3-CAR T cells were stimulated once (day 2) or three times (day 9) with HCC cells at a 1:1 ratio, and rate of apoptosis was evaluated by staining for annexin V. Representative flow plots and summary bar graph for indicated CAR T-cell groups (mean ± SEM, combined data from four independent experiments evaluating 4 donors). Data in B and C were analyzed using two-way ANOVA. *, *P < 0.05; **, **P < 0.01; ***, ***P < 0.001.
Figure 4.
Coexpression of IL21 and IL15 altered global gene-expression patterns in GPC3-CAR T cells, and TCF-1 was maintained in CAR T cells coexpressing both IL15 and IL21.

A, Heat maps showing fold expression changes for top 20 genes with highest or lowest change in expression and reaching significance versus GBBz T cells (arranged with respect to 21.15.GBBz vs. GBBz), as measured 3 days after stimulation with HCC cells (four independent cocultures evaluating 4 independent donors; Nanostrings performed as a single batch).

B–D, TCF-1 protein expression within CD4\(^+\) and CD8\(^+\) GPC3-CAR T cells as measured by intracellular flow cytometry. A representative histogram (B) and combined results showing percentage (C) and mean fluorescence intensity (MFI; D) of TCF-1\(^+\) cells (mean ± SD; combined data from four independent experiments evaluating 4 independent donors). Two-way ANOVA: *, *P < 0.05; **, **P < 0.01; ***, **P < 0.001.

E, Expansion of indicated GPC3-CAR T-cell populations with or without TCF-1 KO after stimulation with GPC3\(^+\) tumor cells (mean ± SD; combined results from three independent experiments evaluating 5 independent donors; Student t test: P < 0.001).

combination improved TCF-1 protein expression in both CD4+ and CD8+ CAR T cells (Fig. 4B and C). After two consecutive stimulations with tumor cells, the percentage of TCF-1–positive cells was the highest in 21.15.GBBz T-cell group for the CD4 subset (21.15.GBBz vs. 21.15.GBBz, $P = 0.002$; 15.GBBz vs. 21.15.GBBz, $P = 0.0296$) whereas in the CD8 subset, the 15.GBBz and 21.15.GBBz T cells had the highest percentage of TCF-1–positive cells (15.GBBz vs. 21.15.GBBz $P = 0.0124$; 21.15.GBBz vs. 21.15.GBBz $P < 0.001$; Fig. 4C). TCF-1 expression in the CD8 subset remained highest after two stimulations in the 21.15.GBBz group compared with all other groups (21.GBBz vs. 21.15.GBBz, $P = 0.0044$; Fig. 4D).

To determine whether TCF-1 was required for the enhanced proliferative capacity of 21.15.GBBz T cells, we knocked out TCF7 with CRISPR-Cas9 and measured CAR T-cell expansion upon repeated engagement with GPC3+–positive tumor cells (Supplementary Fig. S9). We found that TCF7 KO eliminated the proliferative benefit of 21.15.GBBz T cells compared with GBBz T cells (Fig. 4E). These results suggested that IL21- and IL15–mediated maintenance of TCF-1–positive cells was responsible for the increased proliferative capacity of 21.15.GBBz T cells.

**Coexpression of IL15 and IL21 enhanced** in vivo **expansion, persistence, and antitumor activity of GPC3-CAR T cells**

To evaluate the in vivo expansion and persistence of GPC3-CAR T cells, we established HCC xenografts in NSG mice and injected T cells cotransduced with the individual GPC3-CAR constructs and an eGFP. Fluc construct optimized for tracking small numbers of cells in vivo via bioluminescence imaging (Fig. 5A; ref. 32). As observed in our previous study (27), GBBz T cells expanded effectively for 8 days, after which the population contracted and disappeared entirely by 15 days after injection (Fig. 5B and C; ref. 27). 21.15.GBBz and 15.GBBz T cells had a similar timeline of peak expansion compared with GBBz T cells but persisted longer in vivo before their numbers began to decline (day 12, $P < 0.001$). Coexpression of IL15 and IL21 induced the highest expansion and sustained persistence of GPC3-CAR T cells in vivo (day 15, 21.15.GBBz vs. 21.15.GBBz $P = 0.0012$). Peripheral blood analysis 15 days after adoptive transfer showed increased frequency of CD8+ 15.GBBz and 21.15.GBBz T cells compared with GBBz T cells ($P = 0.0076$ and $P < 0.0002$, respectively; Supplementary Fig. S10). In the spleen, the frequency of CD4+ 21.15.GBBz and 15.GBBz CAR T cells was significantly elevated compared with GBBz T cells ($P = 0.0386$ and $P = 0.003$, respectively). The frequency of CD8+ 21.15.GBBz cells was significantly higher in the spleen compared with other groups including 15.GBBz ($P = 0.0083$; Fig. 5D).

As a safety assessment, serum concentrations of IL15 and IL21 were measured in all therapeutic groups (day 15). IL21 and IL15 serum concentrations in mice treated with cytokine-containing CAR T cells measured in all therapeutic groups (day 15). IL21 and IL15 coexpressing CAR T-cell groups induced a more rapid antitumor effect (week 5: 21.15.GBBz/15.GBBz vs. 21.GBBz, $P < 0.001$; Supplementary Fig. S12). In this model, 21.15.GBBz T cells eliminated tumors more rapidly than 15.GBBz T cells (week 4, $P < 0.001$). Lastly, to further stress the antitumor potential of GPC3-CAR T cells, we injected a low dose of 5 × 10⁶ CAR T cells in mice engrafted with rapidly growing HCC xenografts. At this dose, only 21.15.GBBz T cells, but not 15.GBBz T cells, maintained antitumor activity, which translated into significant survival advantage (15.GBBz vs. 21.15.GBBz, $P < 0.001$, Fig. 5E–G). These results demonstrated that 21.15.GBBz T cells had superior expansion, persistence, and antitumor activity against HCC in vivo.

**Discussion**

Here, we showed preclinical evidence that human GPC3-CAR T cells coexpressing IL15 and IL21 had superior expansion and antitumor activity in preclinical models of HCC. Gene-expression analysis identified TCF-1 as a key transcription factor associated with the increased proliferative capacity of 21.15.GBBz T cells.

We found that coexpression of IL21 and/or IL15 with GBBz did not affect the potent, specific, short-term in vitro cytolytic activity of T cells against HCC tumors; in contrast, we did detect significant differences in effector cytokine production polarization when comparing GBBz T cells with those coexpressing one or both cytokines. As in our previous study, GBBz T cells secreted a Th1-polarized cytokine profile (high IFNγ and GM-CSF; low IL10 and IL4; ref. 27). This overall trend was recapitulated in the current study, with CAR T-cell groups showing GBBz-mediated Th1 polarization regardless of cytokine coexpression. However, we found a striking decrease in IL13 production in CAR T cells coexpressing IL15 (15.GBBz and 21.15.GBBz T cells). IL13 is a Th2 cytokine primarily produced by CD4+ T cells that generates many of the same biological effects as IL4, including decreasing the antitumor function of T cells and promoting tumor cell proliferation (40, 41). IL13 also plays an important role in homeostasis of myeloid-derived suppressor cells (MDSC), which can dampen the efficacy of immunotherapies, induce metastasis formation, cancer progression and inhibit CAR T-cell activity (41–45); therefore, limiting the amount of IL13 in the tumor microenvironment may enhance the therapeutic potential of CAR T cells (45, 46).

Because expansion and persistence of CAR T cells are strong predictors of clinical activity, a key objective of this study was to improve these parameters. We demonstrated that coexpression of IL15 and IL21 in GPC3-CAR T cells increased the proportion of naïve/stem cell memory and central memory GPC3-CAR T cells after manufacture. These less-differentiated T cells have greater proliferative capacity than more mature cells (47–49), providing a potential proliferative advantage for GPC3-CAR T cells coexpressing IL12. Given that all experimental groups were manufactured under the same culture conditions including supplementation with IL15 and IL21, this finding was unexpected. Continuous production of IL21 via transgenic expression from the GPC3-CAR throughout the culturing process likely influenced the T-cell phenotype.
Figure 5.
Coexpression of IL15 and IL21 enhanced in vivo expansion, persistence, and antitumor activity of GPC3-CAR T cells. A, Schematic of in vivo evaluation scheme for GPC3-CAR T-cell persistence. NSG mice were injected with $2 \times 10^6$ Huh-7 cells, followed by $2 \times 10^6$ Fluc+ CAR T cells 2 weeks later. B, Monitoring of bioluminescent GPC3-CAR T cells at indicated time points after injection. C, GPC3-CAR T-cell bioluminescence counts (mean ± SEM) over experimental time course. D, Ratio of CD4+ and CD8+ GPC3-CAR T cells relative to mouse CD45-expressing cells in splenic tissue on day 18 as measured by flow cytometry (mean ± SD, one representative of two independent experiments, n = 8–10/GPC3-CAR T group). E, Schematic of in vivo evaluation scheme for GPC3-CAR T-cell antitumor activity. NSG mice were injected with $2 \times 10^6$ Huh-7/Fluc cells, followed by $0.5 \times 10^6$ CAR T cells on day 7. F, Weekly monitoring of bioluminescent Huh-7 tumor cells. G, Kaplan-Meier survival analysis of tumor-bearing mice pictured in F. Data in C and D were analyzed using one-way ANOVA. Combined results from two independent experiments of 11 to 14 animals per GPC3-CAR T-cell group. Survival was estimated by the Kaplan-Meier method and compared by the Gehan-Breslow-Wilcoxon test. *P < 0.05; **P < 0.01; ***P < 0.001.
We determined that compared with other experimental groups, coexpression of IL15 alone or in combination with IL21 decreased the proportion of transduced T cells undergoing apoptosis after multiple stimulations with HCC tumor cells; thus, proportionately increasing surviving CAR T cells. Finally, we found that the TCF7 gene encoding TCF-1, a key transcription factor in T-cell development, expansion, memory formation, and survival (38, 39, 50, 51), was more highly expressed in the CD8+ subsets of GPC3-CAR T cells expressing IL15, IL21, or both compared with in CD8+ GBt2 T cells. Following two rounds of stimulation with HCC cells, TCF-1 expression was highly maintained in 21.15.GBt2 T cells and was associated with enrichment for and continued expansion of CD8+ CAR T cells. CRISPR-Cas9 knockout of TCF7 eliminated the proliferative advantage of 21.15.GBt2 T cells, suggesting that TCF-1 plays a role in enhancing the expansion and survival of IL15- and IL21 coexpressing GPC3-CAR T cells.

In cancer patients, the effector-to-target ratio of CAR T cells to cancer cells greatly favors cancer; thus, its complete elimination requires CAR T cells to kill repeatedly and expand. In sequential killing assays, in which GPC3-CAR T cells are repeatedly exposed to fresh tumor cells in vitro, we found that combined expression of IL15 and IL21 resulted in the most expansion. Additionally, in an aggressive xenograft model of HCC treated with the lowest dose of GPC3-CAR T cells, thereby stressing their functional capacity, cells transduced with 21.15.GBt2 resulted in significant survival advantage. These findings suggest that IL15 and IL21 should provide the most potent antitumor activity in the clinical setting.

Safety remains a central requirement for all cancer treatments. Elevated concentrations of IL15 and IL21 can cause side effects as described in early-phase studies of subcutaneous or intravenous recombinant IL21 and IL15 administration in patients with cancer (52). In these studies, at the maximum tolerated dose (MTD), peripheral blood peak concentrations of IL15 and IL21 were 1.608 pg/mL and 141 ng/mL, respectively (53, 54). In mice infused with GPC3-CAR T cells expressing IL15 and/or IL21, cytokine concentrations at the peak of T-cell expansion remained 100- to 1000-fold below the peak corresponding MTDs measured in humans. Therefore, we do not anticipate systemic toxicities in patients treated with GPC3-CAR T cells coexpressing IL15 and/or IL21. Nevertheless, we showed that the use of inducible caspase-9 can effectively eliminate IL15 and IL21 expressing GPC3-CAR T cells in vivo; thus, this “suicide switch” system may be used during clinical evaluation (33).

In conclusion, GPC3-CAR T cells coexpressing IL15 and IL21 were effective at treating HCC in preclinical models. Our findings address a major barrier by enhancing the expansion and persistence of therapeutic cells, resulting robust antitumor responses. We provided mechanistic insight and preclinical evidence to substantiate testing GPC3-CAR T cells coexpressing IL15 and IL21 in patients, an avenue that will be explored in two ongoing clinical trials (NCT02932956 and NCT02905188).

Disclosure of Potential Conflicts of Interest

S.A. Batra, P. Rathi, L.S. Metelitsa, and A. Heczey have pending patent applications related to GPC3-CARs and/or cytokine coexpression. No potential conflicts of interest were disclosed by the other authors.

Authors’ Contributions

Conception and design: S.A. Batra, P. Rathi, A. Heczey Development of methodology: S.A. Batra, P. Rathi, T.P. Nguyen, M. Mamonkin, L.S. Metelitsa, A. Heczey Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): S.A. Batra, L. Guo, J. Fleurence, R.S. Shaik, T.P. Nguyen, A. Heczey Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): S.A. Batra, P. Rathi, A.N. Courteny, J. Fleurence, J. Baleau, R.S. Shaik, M.-F. Wu, S. Bulsara, A. Heczey Writing, review, and/or revision of the manuscript: S.A. Batra, P. Rathi, A.N. Courteny, J. Fleurence, R.S. Shaik, M.-F. Wu, S. Bulsara, L.S. Metelitsa, A. Heczey Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): S.A. Batra, P. Rathi, L. Guo, A.N. Courteny, J. Fleurence, J. Baleau, R.S. Shaik, T.P. Nguyen, M.-F. Wu, S. Bulsara, M. Mamonkin, L.S. Metelitsa, A. Heczey Study supervision: A. Heczey

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References


IL15 and IL21 Armored GPC3-CAR T Cells against Liver Cancer

A review of potential conflicts of interest

S.A. Batra, P. Rathi, L.S. Metelitsa, and A. Heczey have pending patent applications related to GPC3-CARS and/or cytokine coexpression. No potential conflicts of interest were disclosed by the other authors.

Authors' contributions


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