Human GUCY2C-Targeted Chimeric Antigen Receptor (CAR)-Expressing T Cells Eliminate Colorectal Cancer Metastases

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Abstract

One major hurdle to the success of adoptive T-cell therapy is the identification of antigens that permit effective targeting of tumors in the absence of toxicities to essential organs. Previous work has demonstrated that T cells engineered to express chimeric antigen receptors (CAR-T cells) targeting the murine homolog of the colorectal cancer antigen GUCY2C treat established colorectal cancer metastases, without toxicity to the normal GUCY2C-expressing intestinal epithelium, reflecting structural compartmentalization of endogenous GUCY2C to apical membranes comprising the intestinal lumen. Here, we examined the utility of a human-specific, GUCY2C-directed single-chain variable fragment as the basis for a CAR construct targeting human GUCY2C-expressing metastases. Human GUCY2C-targeted murine CAR-T cells promoted antigen-dependent T-cell activation quantified by activation marker upregulation, cytokine production, and killing of GUCY2C-expressing, but not GUCY2C-deficient, cancer cells in vitro. GUCY2C CAR-T cells provided long-term protection against lung metastases of murine colorectal cancer cells engineered to express human GUCY2C in a syngeneic mouse model. GUCY2C murine CAR-T cells recognized and killed human colorectal cancer cells endogenously expressing GUCY2C, providing durable survival in a human xenograft model in immunodeficient mice. Thus, we have identified a human GUCY2C-specific CAR-T cell therapy approach that may be developed for the treatment of GUCY2C-expressing metastatic colorectal cancer. Cancer Immunol Res; 6(5): 509–16. ©2018 AACR.

Introduction

Chimeric antigen receptor (CAR)-expressing T cells are effective in patients with advanced B-cell malignancies and neuroblastoma (1–3). However, the clinical utility of adoptively transferred T cells has been limited in patients with colorectal cancer, reflecting antigen “on-target, off-tumor” toxicities limiting treatment options (4, 5). Guanylyl cyclase C (GUCY2C) is a membrane-bound receptor that produces the second messenger cGMP following activation by its hormone ligands guanylin or uroguanylin, regulating intestinal homeostasis, tumorigenesis, and obesity (6). GUCY2C cell surface expression is confined to luminal surfaces of the intestinal epithelium and a subset of hypothalamic neurons (7). Its expression is maintained in >95% of colorectal cancer metastases, and it is ectopically expressed in tumors that evolve from intestinal metaplasia, including esophageal, gastric, and pancreatic cancers (8, 9). The inaccessibility of GUCY2C in the apical membranes of polarized epithelial tissue (10), due to subcellular restriction of GUCY2C, creates a therapeutic opportunity to target metastatic lesions of colorectal origin that have lost apical–basolateral polarization (11), without concomitant intestinal toxicity. Previously, we demonstrated in a syngeneic, immunocompetent mouse model that CAR-T cells targeting murine GUCY2C were effective against colorectal cancer metastatic to lung in the absence of intestinal toxicities (12). Similarly, other GUCY2C-targeted therapeutics, including antibody–drug conjugates (13) and vaccines (14, 15), are safe in preclinical animal models, and therapeutic regimens utilizing these platforms are in clinical trials for metastatic esophageal, gastric, pancreatic, and colorectal cancers (NCT02202759, NCT02202785, and NCT01972737). The safety of these therapeutic regimens, in the context of GUCY2C expression across the rostral–caudal axis of intestine, reflects compartmentalized expression of GUCY2C, enriched in apical, but limited in basolateral, membranes of epithelial cells (16–19). Systemic radioiodinated imaging agents conjugated to GUCY2C ligand target GUCY2C-expressing metastases without localizing in intestine (10), confirming the mucosal compartmentalization of the receptor. Tumors express up to 10-fold greater amounts of GUCY2C, compared with normal epithelial cells (9, 20), potentially creating a quantitative therapeutic window to discriminate receptor overexpressing tumors from intestinal epithelium (21) with low/absent GUCY2C in basolateral membranes (19).
Here, we identified a human GUCY2C-targeted CAR that could potentially be used in patients with GUCY2C-expressing gastrointestinal malignancies. This CAR induced antigen-dependent T-cell activation, cytokine production, and cytolytic activity. Human GUCY2C-targeted CAR-T cells were effective against metastatic tumors in immunocompetent, syngeneic mouse models, as well as xenograft models of human colorectal cancer.

**Materials and Methods**

**Cell lines and reagents**

CT26 and β-galactosidase–expressing CT26.CL26 mouse colorectal cancer cell lines and the human colorectal cancer cell lines T84 and SW480 were obtained from ATCC, and large stocks of low-passage cells were cryopreserved. Cells were authenticated by the original suppliers and routinely authenticated by morphology, growth, antibiotic resistance (where appropriate), appropriate GUCY2C and β-galactosidase expression, and pattern of metastasis in vivo and routinely screened for mycoplasma using the Universal Mycoplasma Detection Kit (ATCC, Cat. No. 30-1012K). Before injection into mice, cells were routinely cultured for <2 weeks. The gene encoding human GUCY2C was codon-optimized (Supplementary Fig. S1) and synthesized (Gene Art, Life Technologies) and cloned into the retroviral construct pMSCVpuro (Clontech). CT26.hGUCY2C and CT26.CL25.hGUCY2C were generated by transducing CT26 and CT26.CL25 cells with retroviral supernatants encoding hGUCY2C followed by selection with puromycin. Retroviral supernatants were produced by transfecting the Phoenix-Eco retroviral packaging cell line (Gary Nolan, Stanford University) with pMSCVpuro (Clontech) or hGUCY2C-pMSCV-puro and the pCL-Eco (Imgenex) retroviral packaging vector (12). Luciferase-containing T84.luc cells were generated by transduction with lentiviral supernatants generated by transfecting 293FT cells (Invitrogen) with pLenti4-V5-GW-luciferase.puro (kindly provided by Andrew Aplin, Thomas Jefferson University) and the ViraPower Lentiviral Packaging Mix (Invitrogen) according to the manufacturer’s instructions, followed by selection in puromycin. The single-chain variable fragment (scFv) from the human GUCY2C-specific antibody 5F9 (Supplementary Fig. S2) was cloned into the pPDSE-Rlbg-Fc2 (II2sg) plasmid (InvivoGen), producing a 5F9 scFv fusion protein with rabbit Fc (5F9-rFc). 5F9-rFc was collected in rIgG-Fc2 (IL2ss) plasmid (Invivogen), producing a 5F9 scFv antibody 5F9 (Supplementary Fig. S2) was cloned into the pFUSE-Rev-Eco retroviral vector (Addgene # 27490). The Phoenix-Eco retroviral packaging cell line (Gary Nolan, Stanford University) was transfected with CAR-pMIG vectors and the pCL-Eco retroviral packaging vector (Imgenex) using the Calcium Phosphate Precipitation Mammalian Transfection System (Promega). Retrovirus-containing supernatants were collected 48 hours later, filtered through 0.45 μm filters, and aliquots were frozen at ~80°C.

Murine CD8+ T cells were negatively selected from BALB/c splenocytes using the CD8a+ T-cell Isolation Kit II and LS magnetic columns (Miltenyi Biotec). CD8+ T cells were subsequently stimulated with anti-CD3/anti-CD28–coated beads (T Cell Activation/Expansion Kit, Miltenyi Biotec) at a 1:1 bead:cell ratio at 1 x 10^6 cells/mL in cRPMI with recombinant human IL2 (100 U/mL; NCI Repository). The day following stimulation, half of the culture media were carefully replaced with an equal volume of thawed retroviral supernatant in the presence of polybrene (8 μg/mL, Millipore). Spinning was performed at room temperature for 90 minutes at 2,500 rpm followed by incubation at 37°C for 2.5 hours, at which point cells were pelleted and resuspended in fresh media containing IL2 (100 U/mL). T cells were expanded for 7 to 10 days by daily dilution to 1 x 10^6 cells/mL with fresh cRPMI and IL2, at which point they were used for functional assays.

**Human CAR-T cell generation**

For studies with human T cells, PBMCs were collected from consenting volunteers in accordance with regulatory and institutional requirements. MACS (STEMCELL Technologies) purified CD8+ T cells were negatively selected from individual normal healthy donor whole blood at >90% purity. CAR domains using human sequences were used to produce a third-generation, codon-optimized retroviral CAR construct containing the 5F9 human GUCY2C-specific scFv and human sequences of the GM-CSF signal peptide, CD8a hinge region, CD28 transmembrane activation domain, and 4-1BB (CD137) and CD3ζ intracellular domains producing 5F9.h28BBz CAR construct (Supplementary Fig. S1). CARs derived from the human ERBB2 (Her2)-specific antibody 4D5 or mouse CD19-specific antibody 1D3 (Supplementary Fig. S2) were used as controls as indicated (Control m28BBz). CARs were subcloned into the pMSCV-IRE5-GFP (pMIG) retroviral vector (Addgene # 27490). The Phoenix-Eco retroviral packaging cell line (Gary Nolan, Stanford University) was transfected with CAR-pMIG vectors and the pCL-Eco retroviral packaging vector (Imgenex) using the Calcium Phosphate Precipitation Mammalian Transfection System (Promega). Retrovirus-containing supernatants were collected 48 hours later, filtered through 0.45 μm filters, and aliquots were frozen at ~80°C.

Murine CAR-T cells were stained with the LIVE/DEAD Fixable Aqua Dead Cell Stain Kit (Invitrogen) in PBS, labeled with varying concentrations of 6xHis-hGUCY2C (Addgene # 27490). The Phoenix-Eco retroviral packaging cell line (Gary Nolan, Stanford University) was transfected with CAR-pMIG vectors and the pCL-Eco retroviral packaging vector (Imgenex) using the Calcium Phosphate Precipitation Mammalian Transfection System (Promega). Retrovirus-containing supernatants were collected 48 hours later, filtered through 0.45 μm filters, and aliquots were frozen at ~80°C.

**CAR surface detection**

CAR-transduced T cells were stained with the LIVE/DEAD Fixable Aqua Dead Cell Stain Kit (Invitrogen) in PBS, labeled with varying concentrations of 6xHis-hGUCY2C (Addgene # 27490) for 1 hour in
PBS 0.5% BSA, stained with anti-5xHis–Alexa Fluor 647 conjugate (Qiagen) and anti-CD8b–PE (clone H35.17.2, BD Biosciences) for 1 hour in PBS 0.5% BSA, fixed with 2% PFA, and analyzed using the BD LSR II flow cytometer and FlowJo software v10 (TreeStar). 

Mouse T-cell phenotyping, activation markers, and intracellular cytokine staining

For phenotyping, 1 × 10⁶ nontransduced or CAR-transduced mouse T cells were stained with the LIVE/DEAD Fixable Aqua Dead Cell Stain Kit (Invitrogen) in PBS and subsequently stained for surface markers using anti-CD8α–BV570 (clone RPA-T8; BioLegend), anti-CD45RA–PerCP-Cy5.5 (clone 14.A8; BD Biosciences), and anti-CD62L–PE-Cy7 (clone MEL-14; BD Biosciences) for 30 minutes in PBS 0.5% BSA. Cells were subsequently fixed and permeabilized (BD Cytofix/Cytoperm Kit; BD Biosciences) with Cytofix/Cytoperm buffer for 20 minutes at 4°C and stained for intracellular GFP (anti-GFP–Alexa Fluor 488; Invitrogen) for 45 minutes in Perm/Wash buffer to identify CAR-transduced cells. The following subsets were then quantified based on CD45RA and CD62L staining: Tnexm (naïve or T memory stem cells; CD62L−CD45RA+), Tmn (central memory T cells; CD62L+CD45RA−), Tem (effector memory T cells; CD62L−CD45RA−), and Tmem (effector memory T cells expressing CD45RA−CD62L+CD45RA+).

For intracellular cytokine analysis, 1 × 10⁶ CAR-transduced mouse T cells were stimulated for 6 hours in tissue culture plates previously coated with 1 μg/mL GUCY2C in PBS overnight at 4°C or in tissue culture plates containing 1 × 10⁶ CT26 or CT26.hGUCY2C cells. As a positive control, CAR-T cells were incubated for 6 hours with 1 × 10⁶ CT26 or CT26.hGUCY2C or 2.5 × 10⁶ T84 or SW480 cancer cell targets plated in 150 μL of growth medium in each well of an E-Plate 16 (Acea Biosciences) and grown overnight at 37°C incubator, quantifying electrical impedance every 15 minutes using the RTCA DP Analyzer system and RTCA software version 2.0 (Acea Biosciences Inc.). Approximately 24 hours later for mouse and 6 hours for human T-cell experiments, 50 μL of CAR-T cells were added (5:1 E:T ratio for mouse T cells or 10:1 E:T ratio for human T cells), or 50 μL of media or 10% Triton-X 100 (Fisher) was added for a final (v/v) of 2.5% Triton-X 100 as negative and positive controls, respectively. Cell-mediated killing was quantified over the next 10 to 20 hours, reading electrical impedance every 15 minutes. Percent specific lysis values were calculated using GraphPad Prism Software v6 for each replicate at each time point, using impedance values following the addition of media and Triton-X 100 for normalization (0% and 100% specific lysis, respectively).

Alternatively, the β-gal release T-cell cytotoxicity assay utilized CT26 cancer cell targets expressing β-galactosidase (CT26.CL25). Cancer cell targets were plated at 2 × 10⁵ cells/well in a 96-well plate and incubated with increasing effector CAR-T cell to cancer cell target ratios for 4 hours at 37°C. Released β-galactosidase was measured in the media using the Galacto-Light Plus System (Applied Biosystems). Maximum release was determined from supernatants of cells that were lysed with supplied lysis buffer. % specific lysis = [(experimental release – spontaneous release)/ (maximum release – spontaneous release)] × 100.

Metastatic tumor models

BALB/c mice and NSG (NOD-scid IL2Rγnull) mice were obtained from the NCI Animal Production Program and The Jackson Laboratory, respectively. Animal protocols were approved by the Thomas Jefferson University Institutional Animal Care and Use Committee. In syngeneic mouse models, BALB/c mice were injected with 5 × 10⁵ CT26.hGUCY2C cells in 100 μL of PBS by tail-vein injection to establish lung metastases. On indicated days, mice received a nonmyeloablative dose of 5 Gy total body irradiation in a PanTak, 310 keV X-ray machine (12). Mice were killed, the indicated dose of CAR-T cells produced from CD8⁺ BALB/c T cells in 100 μL of PBS by tail vein at the indicated time points. Mice were followed for survival or sacrificed on day 18 after cancer cell injection and lungs were stained with India Ink and fixed in Fekete’s solution for tumor enumeration (22).
For rechallenge experiments, naïve mice or mice cleared of established tumors by CAR-T cells (referred to as "surviving mice") received one dose of $5 \times 10^5$ CT26 or CT26.hGUCY2C via tail-vein injection. Surviving mice were initially challenged 16 to 40 weeks prior to the rechallenge experiment.

In human tumor xenograft models, NSG (23) mice (JAX stock #005557) were injected with $2.5 \times 10^6$ T84.fLuc cells in 100 µL PBS via intraperitoneal injection. Mice received a dose of $10^7$ total (not sorted on CAR$^+$) T cells produced from CD8$^+$ BALB/c T cells in 100 µL PBS via intraperitoneal injection on day 14 after cancer cell inoculation. Tumor growth was monitored weekly by subcutaneous injection of a 250 µL solution of 15 mg/mL o-luciferin potassium salt (Gold Biotechnologies) in PBS and imaging after 8 minutes of exposure using the Caliper IVIS Lumina-XR imaging station (Perkin Elmer). Total radiance (photons/second) was quantified using Living Image In Vivo Imaging Software (Perkin Elmer).

### Results and Discussion

**hGUCY2C CAR-T cells**

A recombinant antibody (clone 5F9) specific for human GUCY2C (hGUCY2C) bound to purified hGUCY2C extracellular domain (Fig. 1A) and murine CT26 colorectal cancer cells engineered to express hGUCY2C, but not hGUCY2C-deficient CT26 cancer cells (Fig. 1B). The 5F9 scFv was used to generate...
a third-generation murine CAR construct (5F9.m28BBz) containing the BiP signal sequence, CD8α hinge region, and intracellular CD28, 4-1BB, and CD3ζ signaling moieties and inserted into a retroviral construct (Fig. 1C). Retroviruses encoding control m28BBz or 5F9.m28BBz CARs were used to transduce murine T cells with ~35% to 45% transduction efficiency, quantified by a GFP transduction marker (Fig. 1D). hGUCY2C-binding avidity (K_{av} = 11.2 nmol/L) and CAR expression (B_{max} = 957.8 MFI), quantified by incubating CAR-T cells with increasing concentrations of purified 6×His-tagged hGUCY2C_{CExD} followed by detection with labeled 6×His antibody and assessment by flow cytometry, was comparable with CARs that exhibited functional reactivity to mouse GUCY2C (ref. 12; Fig. 1D and E; Supplementary Fig. S5).

hGUCY2C CAR mediates T-cell activation and effector function

Transduction of purified mouse CD8+ T cells with control m28BBz or hGUCY2C-specific 5F9.m28BBz CAR constructs had no impact on T-cell phenotype compared with non-transduced cells (Fig. 2B), producing a mixture of memory and effector phenotypes (T_{HSC}, T_{EM}, and T_{EMRA}) similar to other CAR constructs in CAR-T cells produced in the presence of IL2 (24). hGUCY2C-specific, but not control, CAR-T cells upregulated the activation markers CD25, CD69, and CD44 (Fig. 2C) and produced the effector cytokines IFNγ, TNFα, IL2, and MIP1α (Fig. 2D) when stimulated with immobilized hGUCY2C_{CExD} protein or CT26 hGUCY2C cells (Supplementary Figs. S6 and S7). Activation marker and cytokine responses were absent when 5F9.m28BBz CAR-T cells were stimulated with BSA or hGUCY2C-deficient CT26 cells, confirming that T-cell activation by the 5F9.m28BBz CAR is antigen-dependent (Fig. 2C and D; Supplementary Figs. S6 and S7). Although 5F9.m28BBz CAR-T cells were inactive against hGUCY2C-deficient CT26 cells in vivo (Fig. 2E), they exhibited time-dependent killing of CT26.hGUCY2C cells, quantified using an electrical impedance assay (Fig. 2E) and confirmed in a β-galactosidase release T-cell cytotoxicity assay (Supplementary Fig. S8).

hGUCY2C CAR-T cells oppose metastatic colorectal cancer

The endogenous immune system can induce immunosuppression in the tumor microenvironment and compete with adoptively transferred T cells for resources necessary for long-term persistence (25). In that context, lymphodepleting conditioning regimens, such as low-dose total body irradiation (TBI)


Figure 3.

hGUCY2C CAR-T cells provide long-term protection in a syngeneic lung metastasis model. A-E, BALB/c mice were injected with $5 \times 10^5$ CT26.hGUCY2C cells via the tail vein to establish lung metastases. Control (4D5.m28BBz) or 5F9.m28BBz CAR constructs were transduced into murine CD8+ T cells. A, Mice were treated 3 days later with 5 Gy total body irradiation (TBI) followed by $10^7$ 5F9.m28BBz (N = 7–8/group) or $10^7$ control (N = 6) CAR-T cells. B, Mice were treated on day 3 (D3) or day 7 (D7) with 5 Gy TBI followed by $10^7$ control (N = 10/group) or 5F9.m28BBz (N = 9–10/group) CAR-T cells. C, Mice were treated on day 7 with 5 Gy TBI followed by $10^7$ control (N = 10) or 5F9.m28BBz (N = 12) CAR-T cells on days 7 and 14. D, Mice treated on day 7 with 5 Gy TBI and PBS or $10^7$ control or 5F9.m28BBz CAR-T cells were sacrificed on day 18, lungs stained with India ink, and tumors/lung enumerated. One-way ANOVA; *, P < 0.05; **, P < 0.001; ***, P < 0.0001. Up arrows indicate CAR-T cell treatment days. Each panel indicates an independent experiment.

hGUCY2C CAR-T cells recognize human colorectal tumors

We next determined if hGUCY2C CAR-T cells recognized native hGUCY2C on human colorectal tumors. The recombinant hGUCY2C-specific antibody 5F9 stained hGUCY2C on the surface of GUCY2C-expressing T84 (Fig. 4A), but not GUCY2C-deficient SW480 (Supplementary Fig. S9A) human colorectal cancer cells. Correspondingly, 5F9.m28BBz CAR-T cells lysed T84 (Fig. 4B), but not SW480 (Supplementary Fig. S9B) cancer cells in vitro in a time-dependent manner. Control CAR-T cells did not kill either human cancer cell type, indicating that killing was antigen-dependent (Fig. 4B; Supplementary Fig. S9B). Human T cells expressing a human 5F9 CAR construct (5F9.h28BBz) produced effector cytokines following GUCY2C stimulation and killed human colorectal cancer cells endogenously expressing GUCY2C (Supplementary Fig. S10). Mouse T cells expressing hGUCY2C-specific (5F9.m28BBz), but not control, CAR effectively treated T84 human colorectal tumor xenografts in a peritoneal metastases model (Fig. 4C-E). Together, these data indicated that hGUCY2C-specific CAR constructs produced with the 5F9 scFv can redirect T-cell–mediated killing of human colorectal tumors endogenously expressing hGUCY2C.

Adoptive T-cell therapies targeting colorectal tumor antigens have been limited by antigen “on-target, off-tumor” toxicities (4, 5). We previously validated GUCY2C as a potential target for CAR-T cell treatment in a completely syngeneic mouse model in which CARs targeting mouse GUCY2C promoted
GUCY2C-targeted CAR-T cells are an attractive tool for the T-cell therapy armamentarium, a paradigm that is limited by the lack of suitable antigen targets (30).

Disclosure of Potential Conflicts of Interest
M.S. Magee, T.R. Baybutt, and A.E. Snook have ownership interest in U.S. Provisional Patent Application No. 62/643,850. S.A. Waldman is on the board of directors for Targeted Diagnostics & Therapeutics, Inc., reports receiving a commercial research grant from Targeted Diagnostics & Therapeutics, Inc. and Synergy Pharmaceuticals, Inc., has received speakers bureau honoraria from Synergy Pharmaceuticals, Inc., has ownership interest in Thomas Jefferson University, and is a consultant/advisory board member for Targeted Diagnostics & Therapeutics, Inc. No potential conflicts of interest were disclosed by the other authors.

Figure 4. hGUCY2C CAR-T cells eliminate human colorectal tumor xenografts. A, hGUCY2C expression on T84 human colorectal cancer cells was quantified by flow cytometry using the recombinant 5F9 antibody. B-E, Control (1D3.m28BBz) or 5F9.m28BBz CAR constructs were transduced into murine CDB T cells. B, T84 colorectal cancer cells in an E-Plate were treated in duplicate with 5F9-m28BBz or control CAR-T cells (5:1 E:T ratio), media, or 10% Triton-X 100 (Triton), and the relative electrical impedance was measured every 15 minutes for 20 hours to quantify cancer cell death (normalized to time = 0). Percent specific lysis values were calculated using impendence values following the addition of media and Triton for normalization (0% and 100% specific lysis, respectively). Two-way ANOVA: * *, P < 0.01; representative of two independent experiments. C-E, Immune deficient NSG mice were injected with 2.5 x 10^6 luciferase-expressing T84 colorectal cancer cells via intraperitoneal injection and were treated with PID control (N = 5/group) or 5F9-m28BBz (N = 4/group) CAR-T cells on day 14 by intraperitoneal injection. C and D, Total tumor luminescence (photons/second) was quantified just prior to T-cell injection and weekly thereafter. Two-way ANOVA: * , P < 0.05. E, Mice were followed for survival. Log-rank Mantel-Cox test: * , P < 0.05.

Antitumor efficacy in the absence of toxicities to the normal GUCY2C-expressing intestinal epithelium (12). Here, we produced a human GUCY2C-specific CAR from an antibody that is currently used as an antibody–drug conjugate in clinical trials for GUCY2C-expressing malignancies (NCT02202759 and NCT02202785) and demonstrated its ability to induce T-cell activation, effector function, and antitumor efficacy in both syngeneic and human colorectal tumor xenograft mouse models using murine T cells. CARs produced from the 5F9 scFv do not cross-react with murine GUCY2C (Supplementary Fig. S11), preventing quantification of intestinal toxicity in mouse models. Differences in the antigen-recognition domain of the CAR described here and the murine CAR previously described (12), as well as inherent differences between mice and humans, suggest caution in GUCY2C CAR-T cell administration to humans, despite murine GUCY2C CAR-T cell safety data (12). Thus, appropriate safety measures should be considered when translating the use of GUCY2C CAR-T cells into the clinic, including transient CAR expression by mRNA electroporation (28) or incorporation of suicide genes (29). Nevertheless, GUCY2C-targeted CAR-T cells are an attractive tool for the T-cell therapy armamentarium, a paradigm that is limited by the lack of suitable antigen targets (30).
References


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