Induction of T-cell Immunity Overcomes Complete Resistance to PD-1 and CTLA-4 Blockade and Improves Survival in Pancreatic Carcinoma

Rafael Winograd¹, Katelyn T. Byrne¹, Rebecca A. Evans¹, Pamela M. Odorizzi², Anders R.L. Meyer³, David L. Bajor¹,⁴,⁵, Cynthia Clendenin¹, Ben Z. Stanger¹,⁴,⁵, Emma E. Furth³, E. John Wherry²,⁴, and Robert H. Vonderheide¹,⁴,⁵

Abstract

Disabling the function of immune checkpoint molecules can unlock T-cell immunity against cancer, yet despite remarkable clinical success with monoclonal antibodies (mAb) that block PD-1 or CTLA-4, resistance remains common and essentially unexplained. To date, pancreatic carcinoma is fully refractory to these antibodies. Here, using a genetically engineered mouse model of pancreatic ductal adenocarcinoma in which spontaneous immunity is minimal, we found that PD-L1 is prominent in the tumor microenvironment, a phenotype confirmed in patients; however, tumor PD-L1 was found to be independent of IFNγ in this model. Tumor T cells expressed PD-1 as prominently as T cells from chronically infected mice, but treatment with αPD-1 mAbs, with or without αCTLA-4 mAbs, failed in well-established tumors, recapitulating clinical results. Agonist αCD40 mAbs with chemotherapy induced T-cell immunity and reversed the complete resistance of pancreatic tumors to αPD-1 and αCTLA-4. The combination of αCD40/chemotherapy plus αPD-1 and/or αCTLA-4 induced regression of subcutaneous tumors, improved overall survival, and conferred curative protection from multiple tumor rechallenges, consistent with immune memory not otherwise achievable. Combinatorial treatment nearly doubled survival of mice with spontaneous pancreatic cancers, although no cures were observed. Our findings suggest that in pancreatic carcinoma, a nonimmunogenic tumor, baseline refractoriness to checkpoint inhibitors can be rescued by the priming of a T-cell response with αCD40/chemotherapy. Cancer Immunol Res; 3(4): 399–411. ©2015 AACR.

Introduction

Pancreatic ductal adenocarcinoma (PDA) is a lethal and aggressive disease with the lowest 5-year patient survival rate of any tumor type routinely tracked (6%). The incidence of PDA is rising, and it is projected to become the second leading cause of cancer-related death in the United States by 2025 (1). PDA is distinguished by a dense desmoplastic stroma, rich in fibroblasts, extracellular matrix, and inflammatory leukocytes (but few infiltrating effector T cells). Although new combination chemotherapies are increasingly effective for PDA (2, 3), tumor response rates remain low and durability is short.

T cells are key mediators of antitumor immunity and regulate the outcome of tumor immune surveillance (4). Critical to this regulation are lymphocyte inhibitory receptors, such as programmed cell death protein 1 (PD-1) and cytokotoxic T-lymphocyte–associated antigen 4 (CTLA-4), which restrain T-cell antitumor immunity (5–8). Monoclonal antibodies (mAb) that block PD-1 or CTLA-4 induce T cell–dependent tumor regression in many experimental systems (7, 8). Unprecedented rates of tumor regressions have been observed in patients with melanoma and multiple carcinomas following treatment with mAbs against CTLA-4, PD-1, or programmed death-ligand 1 (PD-L1), a ligand for PD-1 (9–14). PD-1 engagement inhibits T-cell function primarily during the T-cell effector phase (15–17). Its main ligands are PD-L1 and programmed death-ligand 2 (18). PD-L1 is expressed on tumor cells as well as on tumor-associated leukocytes, and antibody blockade of either PD-1 or PD-L1 can restore antitumor immunity in murine cancer models (19–21). CTLA-4 is a crucial immune checkpoint regulator expressed on T cells; loss of CTLA-4 leads to rampant lymphoproliferation, autoimmunity, and death in mice (22). Expression of CTLA-4 is found on effector T cells but especially regulatory T cells (Treg). CTLA-4 blockade in murine tumor models both inhibits negative signaling in effector cells and depletes Tregs (23–25).
Mechanisms of PD-1 or CTLA-4 resistance are poorly understood. Preexisting T-cell antitumor immunity has been hypothesized as a prerequisite (26–29). The majority of cancer patients treated with these agents alone do not respond clinically, and some tumor types, such as PDA, are fully refractory (12, 30, 31). Although combinations of αPD-1 and αCTLA-4 may improve tumor response rates in melanoma, a large fraction of patients still fail to respond (32). Tumor PD-L1 expression in some malignancies correlates spatially with the presence of infiltrating CD8+ T cells, suggesting that tumor cells upregulate PD-L1 in response to immune pressure, a hypothesis termed adaptive immune resistance (33–35). CD8+ T cell–derived IFNγ may drive PD-L1 expression in malignant cells (18, 36). These data suggest that the efficacy of checkpoint inhibitors may require the presence of an endogenous antitumor T-cell response. In fact, the augmentation of antitumor T-cell responses with vaccines, peritumoral IL-2, or oncolytic viruses has been shown to improve baseline responses to checkpoint inhibitors in murine models (21, 27, 37, 38).

In the studies reported here, we tested the hypothesis that failed immune recognition or poor T-cell priming underlies weak clinical responses to checkpoint therapy in pancreatic cancer, that is, induction of T-cell immunity is required to potentiate tumor regressions not otherwise achievable with checkpoint blockade alone. We studied the KPC mouse model of spontaneous PDA in which expression of oncogenic KrasG12D and mutant p53 is targeted to the pancreas by Cre recombinase under the control of the pancreas-specific promoter Pdx-1 (39). This model recapitulates the molecular, histologic, and immune parameters of the human disease (39–43). Analysis of human PDA was performed to confirm the clinical relevance of our findings in the murine model.

We induced T-cell immunity using an agonistic αCD40 in combination with αPD-L1/αCTLA4 mAbs.

Materials and Methods

Mice

All animal protocols were reviewed and approved by the Institutional Animal Care and Use Committee of the University of Pennsylvania. KrasS12G12D/+ Tp53LSL-R172H/+, Pdx1-Cre (KPC) mice (39), and KrasLSL-G12D/+ Tp53LSL-R172H/+, Pdx1-Cre, LSL-Rosa-YFP (KPC-Y) mice (46) were backcrossed 10 generations on the C57BL/6 background. Six- to 8-week-old female C57BL/6 and B6.129S7-Ifngtm1Ts/J (IFNγ ko) mice used for implantable tumor experiments were from The Jackson Laboratory.

Cell lines

PDA cell lines from KPC or KPC-Y mice were derived from single-cell suspensions of PDA tissue as previously described (42). Dissociated cells were cultured in a 6-cm dish with serum-free DMEM. After 2 weeks, media were changed to DMEM + 10% FCS. After 4 to 10 passages, cells were used in experiments. The cell lines were tested and confirmed to be Mycoplasma free. No other authentication assays were performed.

In vivo mouse studies

For implantable tumor experiments, PDA tumor cells (5 × 10^6) were injected subcutaneously in PBS into the flanks of mice and allowed to grow 9 to 11 days until tumor volumes averaged 30 to 100 mm^3. Mice were then assigned to treatment groups such that cohorts were balanced for baseline tumor size. Mice were treated intraperitoneally (i.p.) with αPD-1 (RMp1-14; BioXcell; 200 µg/dose) on days 0, 3, 6, 9, 12, 15, 18, and 21 (after enrollment) and/or αCTLA-4 (9H10; BioXcell; 200 µg/dose) on days 0, 3, and 6. All antibodies were endotoxin free. Clinical grade gemcitabine (Eli Lilly and Company) was purchased through the Hospital of the University of Pennsylvania Clinical; clinical grade nab-paclitaxel was either purchased or a kind gift from Celgene. Chemotherapy vials were resuspended and diluted in sterile PBS, and injected i.p. at 120 mg/kg (for each chemotherapeutic) on day 1. As a control for the human albumin component of nab-paclitaxel, control cohorts were treated with human albumin at the same dose as the albumin component of nab-paclitaxel (108 mg/kg) on day 1 (Sigma Life Science). All antibodies were given i.p. Agonistic αCD40 (FGK45; BioXcell; 100 µg) was given on day 3. For T-cell depletion studies, αCD8 (2.43; BioXcell; 200 µg/dose) and αCD4 mAbs (GK1.5; BioXcell; 200 µg/dose) were injected twice weekly for the duration of the experiment, starting on day 0 (day of enrollment). For isotype controls, rat IgG2a (2A3; BioXcell; 100 µg) and rat IgG2b (LTF-2; BioXcell; 200 µg/dose) were used. This approach achieved >98% depletion of CD8^+ and CD4^+ T cells in peripheral blood and tumor tissue compared with that of control mice, as monitored by flow cytometry. For macophagocyte depletion studies, clodronate-encapsulated liposomes (CEL) or PBS-encapsulated liposomes (PEL; both at 12 µl/g; purchased from Dr. Nico van Rooijen, Vrije Universiteit, Amsterdam, the Netherlands) were used i.p. starting on day –1 and repeated every 4 days for the duration of the experiment; in these experiments, 2.5 × 10^7 PDA cells were implanted. For tumor rechallenge studies, αCD8 or isotype control antibodies were injected i.p. the day before the second rechallenge and continued twice weekly until day 60 or the mouse was sacrificed for tumor burden. To monitor growth of subcutaneous tumors, tumor diameters were measured by calipers and volume calculated by 0.5 × L × W^2 in which L is the longest diameter and W is the perpendicular diameter. Endpoint criteria for the survival studies included tumor volume exceeding 1,000 mm^3 or tumor ulceration. Mice that died suddenly or developed vestibular signs, as described in Supplementary Fig. S8, with minimal tumor burden were censored on the day of death or euthanasia.

For studies using the KPC model, young KPC mice were monitored by abdominal palpation and/or ultrasonography (Vevo 2100 Imaging System with 55MHz MicroScan transducer, Visual Sonics) for the development of pancreatic tumors. Mice with ultrasound diagnosed tumors of volume 30 to 150 mm^3 were enrolled and block randomized into treatment groups. Tumors were visualized and reconstructed for quantification using the integrated Vevo Workstation software package. Baseline tumor volume was not significantly different across cohorts. KPC mice were treated with the same dose and schedule of antibodies and chemotherapeutics as noted above in the subcutaneous model. Mice were censored from study if they developed a secondary malignancy (n = 1). Endpoint criteria included tumor volume exceeding 1,000 mm^3 (by ultrasonography), severe cachexia, or extreme weakness and inactivity.

For viral studies, C57BL/6 mice were infected intravenously with 4 × 10^6 PFU of lymphocytic choriomeningitis viral (LCMV) clone 13, which was propagated, titrated, and used as previously described (15). Mice were sacrificed on day 30 after infection and tissues harvested for analyses.
Collection of tissue samples from mice

The entire pancreas (KPC) or subcutaneous tumor was washed in PBS, minced into small fragments, and incubated in collagenase solution (1 mg/mL collagenase V in DMEM) at 37°C for 45 minutes. Dissociated cells were passed through a 70-μm cell strainer twice and washed three times in DMEM. Spleens and lymph nodes were homogenized and passed through a 70-μm cell strainer to achieve single-cell suspensions. For spleens, red blood cells were lysed using ACK Lysis Buffer (BioWhittaker).

Antibodies, flow cytometry, in vitro IFNγ stimulation of tumor cells, and toxicology are described in Supplementary Methods.

Patient samples and analysis

Formalin-fixed, paraffin-embedded tissue samples were prepared after surgical resection of patients with resectable pancreatic carcinoma according to an Institutional Review Board approved protocol, as noted in Supplementary Methods.

Statistical analysis

Differences between two groups were analyzed by a two-tailed Student t test. Differences between three or more groups were analyzed by one-way ANOVA with the Bonferroni multiple comparison test used as a post hoc test to assess differences between any two groups. Tumor growth curves were analyzed by two-way ANOVA, with Tukey multiple comparisons of means used as a post hoc test to assess differences between any two groups. Survival curves were assessed by the log rank (Mantel–Cox). The correlation between two groups was assessed by the Spearman rank correlation coefficient. All statistical analyses were performed on GraphPad Prism 6 (GraphPad) except two-way ANOVA and related post hoc testing that were performed on R Statistical Software (R Core Team). P ≤ 0.05 denotes differences that are statistically significant.

Results

PD-1–PD-L1 axis is highly expressed in murine and human PDA

We interrogated the expression of PD-1 and PD-L1 using the KPC spontaneous genetic model of PDA. Within the microenvironment of KPC tumors, few infiltrating T cells were observed, as previously reported (41, 42), but these T cells prominently expressed PD-1 in all subsets including CD8⁺, CD4⁺, and regulatory (Foxp3⁺) T cells. For each subset, PD-1 expression was significantly higher in the tumor than in the corresponding populations in the spleens of the same tumor-bearing mice (Fig. 1A). In the absence of a distinct marker for pancreatic epithelial cells in the KPC model, we identified tumor cells with negative gating, excluding leukocytes (CD45), endothelial cells (CD31), and mesenchymal populations (CD90) by flow cytometry of single-cell suspensions of KPC tumors. KPC pancreatic tumor cells exhibited moderate expression of PD-L1 on more than 40% of the identified tumor cells (Fig. 1B). PD-L1 was also expressed by 10% to 50% of normal pancreatic epithelial cells identified in tumor-free wild-type mice. Dendritic cells (DC) and macrophages in the KPC tumor microenvironment expressed very high levels of PD-L1, statistically significantly higher compared with PD-L1 expression of these same antigen-presenting cell (APC) populations in the spleens of KPC mice (Fig. 1B).

To assess whether these findings in the KPC model were consistent with human PDA, we examined human PDA samples for PD-L1 expression. In primary tumors from patients with resected PDA, we observed moderate to intense expression of PD-L1 on tumor and mononuclear cells in 4 of 8 (50%) resection specimens (Fig. 1C and Supplementary Fig. S1). We also observed that T cells in human PDA were relatively rare within malignant foci (mean ratio of CD8⁺ T cells per μm² of tumor vs. non-tumor areas was 0.065 ± 0.052; range, 0.000–0.170)—again consistent with the KPC model (42). PD-L1 expression on tumor cells in human samples did not correspond spatially with the presence of CD8⁺ T cells; there was no statistical correlation between intensity or extent of tumor PD-L1 expression and intratumoral CD8⁺ T-cell infiltration (P = 0.69; Fig. 1C). For example, of the two tumors with the most intratumoral CD8⁺ T cells, one had intense and the other had minimal PD-L1 expression (Fig. 1C). These data in human PDA are in contrast to the correspondence of tumor PD-L1 expression and T-cell infiltration previously reported for tumors from patients with melanoma or kidney or head and neck carcinoma (HNSCC; refs. 33–35).

PD-1 is as highly expressed in murine PDA as it is in chronic LCMV infection

To evaluate the potential role of the PD-1–PD-L1 axis in mediating immune suppression in PDA, we first generated a PDA cell line from a backcrossed KPC mouse and established subcutaneous PDA tumors in immunocompetent C57BL/6 mice. Histopathologic examination of established tumors from this model showed recapitulation of both the cellular and extracellular components of spontaneous KPC tumors, with prominent deposition of a dense desmoplastic stroma and comparable populations of infiltrating immunosuppressive leukocytes, including F4/80⁺ macrophages (data not shown).

We then examined expression of PD-1 on T cells from subcutaneous tumor-bearing mice but did so by simultaneously examining PD-1 expression on T cells from a parallel cohort of mice in which chronic LCMV infection had been established with LCMV clone 13 (Fig. 2A). In many ways, this model of chronic LCMV infection has served as a gold standard for understanding the transcriptional basis and phenotype of exhausted CD4⁺ and CD8⁺ T cells (15, 16, 47–50). The most strongly upregulated genes mechanistically linked to T-cell exhaustion in response to chronic infection in this model are PD-1 and Lag-3 (15). We therefore compared coexpression of these markers on intratumoral and splenic T cells in mice bearing established subcutaneous PDA tumors with splenic T cells from mice with chronic LCMV (Fig. 2A). Intratumoral CD8⁺, CD4⁺, and Tregs coexpressed PD-1 and Lag-3 at levels comparable with the same T-cell populations in LCMV-infected mice (Fig. 2B). This phenotype was restricted to the tumor, as splenic T cells from tumor-bearing mice did not coexpress PD-1 or Lag-3. Thus, T-cell expression of PD-1 is as prominent in the PDA tumor microenvironment as it is in chronic LCMV infection.

In the subcutaneous PDA model, about 60% to 70% of tumor cells isolated from established tumors expressed PD-L1 (Fig. 2C), similar to the expression of PD-L1 on this cell line grown in vitro (Fig. 2D). These findings were confirmed using a YFP⁺ tumor cell line established from a pancreatic tumor isolated from a KPC-Y genetically engineered mouse; in this model, YFP serves as a validated lineage tracer for pancreatic epithelium (46). After subcutaneous tumor implantation and growth in syngeneic hosts, we found that on average 66.7% of YFP⁺ tumors cells expressed PD-L1, as measured by flow cytometry (Supplementary Fig. S2).
Moreover, high levels of PD-L1 on both DCs and macrophages were observed in the tumor microenvironment of the KPC subcutaneous tumors (Fig. 2C), mirroring PD-L1 expression on these APC subsets in spontaneous tumors of KPC mice. Both a higher percentage of PD-L1+ APCs and a higher (~3-4-fold) mean fluorescence intensity (MFI) of PD-L1 were observed compared with that of the corresponding APC populations in the spleens of the same mice (Fig. 2C). These data indicate that PD-L1 expression is prevalent in the PDA microenvironment.

Tumor PD-L1 expression in PDA is not IFNγ dependent

PD-L1 expression in human melanoma and HNSCC correlates spatially with T-cell infiltration (33, 35), and in melanoma, tumor expression of PD-L1 is dynamically upregulated in response to IFNγ secreted by infiltrating CD8+ T cells (36). To determine whether this same mechanism is responsible for PD-L1 expression in PDA, we assessed the ability of our PDA cell line to upregulate PD-L1 in response to IFNγ; in vitro, IFNγ stimulation resulted in increased expression of PD-L1 by PDA cells (Fig. 2D). In vivo, we evaluated tumor and APC PD-L1 expression in the presence or absence of T cells and IFNγ. Subcutaneous PDA tumors were established in mice that were genetically lacking IFNγ; systemically depleted of CD4+ and CD8+ T cells, or both. Tumor growth rate in vivo was the same for each condition compared with that of control (data not shown). Analysis of these tumors showed no significant change in tumor PD-L1 expression (either percentage or MFI) with regard to IFNγ or T-cell status (Fig. 2E). Analysis of the APC populations in these tumors indicated that IFNγ plays a minor role in the regulation of PD-L1 expression on intratumoral DCs and macrophages. Small but statistically significant differences were observed in the percentage and MFI of PD-L1 expression on intratumoral APCs between IFNγ-sufficient and IFNγ-deficient hosts (Fig. 2F). In contrast, the presence or absence of T cells did not affect PD-L1 expression by APCs regardless of host IFNγ status (Fig. 2F), recapitulating the lack of correspondence between CD8+ T cells and PD-L1 expression in human PDA (Fig. 1C).

T-cell stimulation with CD40/gemcitabine/nab-paclitaxel converts PDA from being fully refractory to highly sensitive to checkpoint blockade

We then tested the antitumor in vivo efficacy of PD-1-blocking mAbs either with or without CTLA-4 blocking mAbs (Fig. 3A). CTLA-4 is expressed at higher levels on intratumoral CD8+ and CD4+ T-cell populations (including Treg) when compared with that of splenic populations, suggesting that blockade of this negative checkpoint may improve antitumor immunity (Supplementary Fig. S3). Even with CTLA-4, PD-1 did not affect tumor growth or survival (Fig. 3B), even though a comparable PD-1 dosing schedule reproducibly improves clinical outcomes in mice chronically infected with LCMV clone 13 (50, 51). This lack of antitumor efficacy is similar to the lack of responses observed to date in patients with advanced PDA treated with αPD-L1 or αCTLA-4 (12, 30, 31).

These same reagents have shown efficacy in patients with other malignancies; one possible distinction may be the presence of an antitumor immune response at baseline in subsets of these patients (28, 52). We therefore hypothesized that the induction of a T-cell response would be required to overcome refractoriness to αPD-1 and αCTLA-4 in PDA and achieve clinical benefit. Agonist αCD40 antibody facilitates cancer vaccines (53) and can synergize with chemotherapy-induced immunogenic cell death to initiate a T-cell-dependent antitumor regression, providing a vaccine effect in model systems for which a tumor-rejection antigen is not characterized (45, 54). APCs in KPC-derived subcutaneous tumors express CD40 (Supplementary Fig. S4), as we have previously shown (42). Here, we studied gemcitabine and nab-paclitaxel because this combination was recently approved by the FDA for the treatment of metastatic PDA (3), and gemcitabine has been shown to cooperate immunologically with αCD40 (45). Treatment of mice with established subcutaneous PDA tumors with αCD40/chemotherapy altered the phenotype of tumor-infiltrating T cells although the percentage of T cells infiltrating the tumors did not change. There were statistically significantly fewer CD8+ T cells that coexpressed the inhibitory PD-1 and Lag-3 markers in treated tumors, and more proliferating CD4+ and CD8+ T cells were found in the tumors of treated mice compared with controls (Supplementary Fig. S5).

The combination of gemcitabine/nab-paclitaxel at the maximum-tolerated dose did not induce a rejection of established subcutaneous PDA tumors; however, the addition of αCD40 to this chemotherapy regimen inhibited tumor growth and improved survival compared with control-treated tumor-bearing mice (Fig. 3C). These effects were macrophage independent as CEL failed to abrogate the effect (Supplementary Fig. S6). Given that we have previously observed that αCD40 plus chemotherapy in the subcutaneous KPC model manifests an antitumor effect that is T-cell dependent (55), we added αPD-1, αCTLA-4, or both to treatment with αCD40/chemotherapy in an effort to enhance T-cell immunity. The addition of αPD-1, αCTLA-4, or both to αCD40/chemotherapy enhanced tumor growth inhibition and led to an increase in survival in mice bearing subcutaneous PDA tumors (Fig. 3C). Moreover, these combinations led to complete rejection of established tumors and long-term tumor-free survival in significant proportions of mice treated with the combined regimen (Fig. 3D). Treatment with αPD-1 and αCTLA-4 did not alter CD40 expression on APCs in the tumor microenvironment (Supplementary Fig. S4). The highest rates of tumor regression were observed in mice treated with both αPD-1 and αCTLA-4.
Winograd et al.

A

**C57BL/6**

**Day:** 0 16 30

**Tumor injection**

**Sacrifice**

B

**CD8+ T cells**

**Tumor**

**Spleen**

**Cl-13 spleen**

**CD4+ T cells**

**Regulatory T cells**

C

**Isotype**

**PD-L1**

**Tumor cells**

**Tumor DCs**

**Spleen DCs**

**Tumor Macs**

**Spleen Macs**

D

**Isotype**

**PD-L1**

**PD-L1 (IFN-γ)**

**Tumor cell line**

E

\[
P = 0.076
\]

**B6**

**B6 TCD**

**B6 IFN-γ ko**

**B6 IFN-γ ko TCD**

**P = 0.076**

**B6**

**B6 TCD**

**B6 IFN-γ ko**

**B6 IFN-γ ko TCD**

F

\[
P = 0.58
\]

**B6**

**B6 TCD**

**B6 IFN-γ ko**

**B6 IFN-γ ko TCD**

**P = 0.58**

**B6**

**B6 TCD**

**B6 IFN-γ ko**

**B6 IFN-γ ko TCD**

Winograd et al.

Cancer Immunol Res; 3(4) April 2015

Cancer Immunology Research

Published OnlineFirst February 12, 2015; DOI: 10.1158/2326-6066.CIR-14-0215

Downloaded from cancerimmunolres.aacrjournals.org on June 14, 2021. © 2015 American Association for Cancer Research.
with 39% (17 of 44) of mice achieving long-term complete remission and survival after treatment with all three antibodies plus chemotherapy (Fig. 3D). Tumor growth was delayed in nearly all mice treated with αCD40/chemotherapy and αPD-1/αCTLA-4, even in those mice not completely rejecting their tumors, suggesting that the tumor response rate is even higher than the tumor rejection rate in this model (Supplementary Fig. S7). Although treatment was well tolerated in the vast majority of mice, in 6.3% of mice treated with αCD40/chemotherapy and at least one checkpoint blocking mAb we noted clinical deterioration consistent with an infectious syndrome (Supplementary Fig. S8).

### Rejection of PDA tumors by αCD40/chemotherapy and checkpoint blockade is T cell mediated

To determine whether the antitumor effect we observed was T cell mediated, we repeated the study with a cohort of mice depleted in CD8 T cells, starting on the day before the initiation of therapy. In the absence of T cells, the treatment did not inhibit tumor growth or afford survival advantage, and no T cell–depleted mice rejected the tumor or survived long term (Fig. 4A).

To understand the effect of our treatment on intratumoral T-cell populations, we treated cohorts of tumor-bearing mice with αPD-1/αCTLA-4, αCD40/chemotherapy, both, or neither (control), and sacrificed mice 1 week after treatment with αCD40 or (control) to analyze tumors for T-cell infiltration. Tumors from mice treated with αPD-1/αCTLA-4 plus αCD40/chemotherapy had a significantly increased (7-fold) CD8:Treg ratio compared with that of control-treated mice (Fig. 4B). This phenotype was also seen in some of the mice treated with αCD40/chemotherapy or αPD-1/αCTLA-4, although neither one of these two groups exhibited a relative increase in the CD8:Treg ratio as the mice treated with αPD-1/αCTLA-4 plus αCD40/chemotherapy (Fig. 4B). The CTLA-4 mAb clone 9H10 partly mediates its antitumor effect by depletion of Tregs, which express CTLA-4 (24); however, we observed that mice treated with αPD-1/αCTLA-4 alone did not have a significantly decreased percentage of Tregs among CD4+ T cells (Fig. 4B) or among total CD45+ cells. Rather, the administration of αCD40/chemotherapy (either with or without αPD-1/αCTLA-4) was associated with a significant decrease in Treg percentages compared with that of control-treated mice; all groups treated with immunotherapy demonstrated a slight increase in the CD8+ T-cell infiltrate, although the changes were not statistically significant (Fig. 4B). These data suggest that αCD40/chemotherapy changes the immune microenvironment in this PDA model and leads to a decreased percentage of Tregs and increased CD8:Treg ratio, an effect that is augmented further with the addition of checkpoint blockade. The greatest changes in Treg percentage and CD8:Treg ratio were associated with the highest rates of complete remission and long-term survival across cohorts reported in Fig. 3D.

To test whether mice that had completely rejected established PDA tumors had developed an effective immune memory, we rechallenged cohorts of mice that were in long-term complete remission with the same number of cells of the same PDA tumor line but on the opposite flank (Fig. 4C). We observed that 67% to 86% of such mice rejected the PDA tumor cells implanted on the opposite flank without any additional therapy (Fig. 4C), consistent with those mice having developed immunologic memory. Because the most likely effector memory T-cell population mediating this effect is a CD8+ T cell, we further studied mice that had rejected both the initial tumor and the first rechallenge on the opposite flank, and either depleted these mice of CD8+ T cells or administered an isotype control antibody that did not deplete CD8+ T cells. All mice were then rechallenged with the same number of cells of the same cell line on the original flank. All mice depleted of CD8+ T cells rapidly developed progressively growing tumors at the site of the second rechallenge, whereas 4 of 6 isotype-treated mice rejected this second tumor rechallenge (Fig. 4D). This effect translated into a statistically significant difference in overall survival after the second rechallenge (Fig. 4D).

### αCD40/chemotherapy cooperates with PD-1 blockade to improve survival of mice with established tumors in the KPC genetic model of PDA

Having observed that the induction of T-cell immunity via αCD40/chemotherapy potentiates the efficacy of checkpoint inhibitors in the subcutaneous model of PDA, we then tested this approach in the autologous KPC model of PDA. Observations in the KPC model have previously been shown to predict clinical responses in PDA patients treated with the same or homologous agents (42, 56–58). We therefore performed a randomized, controlled study of checkpoint inhibition in combination with αCD40/chemotherapy in cohorts of tumor-bearing KPC mice. Given the striking expression of PD-1 and PD-L1 in the KPC tumor microenvironment, we chose to test our hypothesis using the αPD-1 mAb. Mice diagnosed with pancreatic tumors of 30 to 150 mm3 were randomized to treatment with αCD40 plus gemcitabine/nab-paclitaxel, αPD-1, αCD40/chemotherapy plus αPD-1, or control (as described in Materials and Methods and summarized in Fig. 5A), using the same dose and schedule as used for mice in the subcutaneous PDA studies. We observed a statistically significant increase in overall survival for mice receiving αCD40/chemotherapy plus αPD-1 compared with that of control (P = 0.015, log-rank Mantel-Cox; Fig. 5B). The effect was large: Combination treatment nearly doubled the median overall survival from 23 days in the control arm to 41.5 days in the
Figure 3.
T-cell stimulation with αCD40/gemcitabine/nab-paclitaxel potentiates efficacy of checkpoint blockade in murine model of PDA. A, experimental design for study of subcutaneous PDA tumors treated with checkpoint inhibitors and αCD40/chemotherapy, as further described in Materials and Methods (G, gemcitabine; nP, nab-paclitaxel; q3d, antibody administered every 3 days). B, tumor growth and survival analyses of mice bearing subcutaneous PDA tumors treated as indicated (n = 9–10 per cohort; results for control and αPD-1 + αCTLA-4 cohorts representative of three independent experiments). See also Supplementary Fig. S7. C, tumor growth and survival analyses of mice bearing subcutaneous PDA tumors treated as indicated (n = 9–10 per cohort; findings representative of three independent experiments). Two-way ANOVA: P \( < 0.0001 \). Post hoc test P values are indicated where statistically significant: *, \( P < 0.05 \); ***, \( P < 0.001 \); ****, \( P < 0.0001 \). See also Supplementary Fig. S7. D, percentage of mice bearing subcutaneous PDA tumors treated with indicated regimens that rejected their tumors and survived tumor-free long-term (median follow-up of 42 days; range, 23–222 days). Data were compiled from five independent experiments.
Figure 4.
Rejection of PDA tumors after αCD40/chemotherapy and checkpoint blockade is T cell-mediated. A, tumor growth and survival analyses of mice bearing subcutaneous PDA tumors treated as indicated (n = 9-10 per cohort; G, gemcitabine; nP, nab-paclitaxel; TCD, CD4/CD8 depletion). B, flow cytometric analysis of subcutaneous PDA tumors treated as indicated (day 18 after tumor injection, day 7 after αCD40 treatment; P, αPD-1; C, αCTLA-4). One-way ANOVASCD8s of live cells: \( P < 0.0001 \); one-way ANOVA%Tregs of CD4+ T cells: \( P = 0.0004 \); one-way ANOVA CD8:Treg ratio: \( P = 0.0005 \). Post hoc test \( P \) values are indicated where statistically significant: \*, \( P < 0.05 \); **, \( P < 0.01 \); ***, \( P < 0.001 \). C, experimental design for first tumor rechallenge experiments. The table quantifies fraction and percentage of mice that rejected tumor rechallenge in mice that had rejected the initial tumor implantation and were tumor free for at least 43 days. Data were compiled from three independent experiments. D, experimental design for second tumor rechallenge experiment. The second rechallenge occurred on days 31 to 49 after the first rechallenge. The table quantifies fraction and percentage of mice that rejected the second tumor rechallenge in mice that had rejected a first tumor rechallenge. Host mice in this experiment were either treated with αCD8 (n = 11) or isotype (Iso; n = 6) antibodies. Survival analysis of mice after the second rechallenge with or without CD8 depletion is shown. Data were compiled from two independent experiments.
Winograd et al.

Discussion

The clinical success of checkpoint inhibitors, including FDA approval of ipilimumab, pembrolizumab, and nivolumab for melanoma, has prompted investigations to replicate these results more broadly in oncology. Early findings, however, suggest that many tumors are resistant, with pancreatic carcinoma appearing completely refractory to monotherapy with checkpoint blockade (12, 30, 31). Here, using a genetically engineered mouse model of PDA, we demonstrate that despite robust expression of PD-1 and PD-L1 in the tumor microenvironment, treatment with αCD40/chemotherapy and αPD-1, as described in Materials and Methods, significantly improved overall survival in KPC genetic model of PDA. A, experimental design for randomized, controlled study in tumor-bearing KPC mice, treated with αCD40/chemotherapy and αPD-1, as indicated in Materials and Methods. G, gemcitabine; nP, nab-paclitaxel; q3d, antibody administered every 3 days. B, overall survival analysis of tumor-bearing KPC mice treated as indicated (n = 6–8 per cohort). αPD-1 alone versus isotype alone P = 0.39; CD40/G/nP vs. isotype alone P = 0.76; CD40/G/nP + αPD-1 vs. isotype alone P = 0.015. C, median overall survival of tumor-bearing KPC mice treated as indicated.

Figure 5. Combination of αCD40/chemotherapy and PD-1 blockade improves survival in KPC genetic model of PDA. A, experimental design for randomized, controlled study in tumor-bearing KPC mice, treated with αCD40/chemotherapy and αPD-1, as described in Materials and Methods. G, gemcitabine; nP, nab-paclitaxel; q3d, antibody administered every 3 days. B, overall survival analysis of tumor-bearing KPC mice treated as indicated (n = 6–8 per cohort). αPD-1 alone versus isotype alone P = 0.39; CD40/G/nP vs. isotype alone P = 0.76; CD40/G/nP + αPD-1 vs. isotype alone P = 0.015. C, median overall survival of tumor-bearing KPC mice treated as indicated.
microenvironment may also be “targetable” as part of novel immunotherapeutic approaches (43, 65, 66).

Despite the 80% increased overall survival observed in KPC mice treated with cetuximab/chemotherapy and αPD-1 compared with that of the controls, all mice succumbed to their disease. It is worth noting that there are no published reports of cures of KPC mice bearing established invasive tumors. A few groups have reported improved overall survival (without cures) with treatment in this model, including the recent demonstration that the vitamin D analogue calcipotriol improves survival by 57% (57, 67, 68). Given the difference we observed in the response to treatment between the subcutaneous and KPC PDA models, we hypothesize that there is additional complexity in the tumor microenvironment of spontaneous KPC tumors that limits therapeutic responses. Potential other immunosuppressive pathways contributing to treatment resistance include myeloid-derived suppressor cells (MDSC), macrophages, and fibroblast-activating protein A (FAP A) mesenchymal cells, among others (43, 65, 66, 69).

Our observations that αCD40/chemotherapy converts PDA from a tumor fully refractory to checkpoint inhibition to one that is sensitive are important in the context of prior efforts to extend the therapeutic range of αPD-1 and αCTLA-4. Our goal was to use a murine model that reproduces the lack of clinical effect observed to date with αPD-1 and αCTLA-4 in PDA, in contrast to many tumor models that exhibit baseline levels of responsiveness to αCTLA-4, αPD-1, αPD-L1, or combinations of these agents. For example, in models of colon carcinoma, melanoma, ovarian cancer, bladder cancer, and neuroblastoma, checkpoint therapy alone inhibits tumor growth, improves survival, and occasionally mediates complete rejection (21, 70–73). In studies using the Panc02 subcutaneous PDA model, tumor rejection was observed in 50% of mice after treatment with αCTLA-4 (74), and treatment with αPD-L1 (with or without ganciclovir) leads to significant decreases in tumor growth (75), findings not representative of the clinical record of these agents in patients with PDA (12, 30, 31) and possibly related to Panc02 being a carcinogen-induced and likely hypermutated tumor [whereas human PDA is not a hypermutated tumor (76)]. Previous work in these types of models demonstrates that T-cell stimulatory therapies can improve baseline effects of checkpoint blockade (21, 27, 37, 38). For example, vaccination of mice bearing metastatic Panc02 tumors with a granulocyte macrophage colony-stimulating factor–secreting tumor cell vaccine was recently described to cooperate with anti–PD-1 therapy, which has single-agent activity in this model (77, 78). In PDA, the stimulation of a T-cell response using αCD40/chemotherapy is able to transform a tumor that is refractory to checkpoint inhibition into a sensitive one, rather than only improving upon baseline activity of checkpoint mAbs.

In the absence of a defined tumor antigen in our KPC model, we therapeutically induced T cells with chemotherapy followed by an agonist αCD40 (45). Although other agents may also synergize with αCD40 (79), gemcitabine, in particular, cooperates with αCD40 (45). Here, we added nab-paclitaxel given the recent regulatory clinical approval of the combination. We have previously demonstrated in KPC mice with spontaneous tumors that αCD40 can reprogram macrophages to mediate (T cell–independent) stromal involution and short-term decreases in tumor volume without improvement in overall survival (42); here, we found that a one-time treatment with chemotherapy followed by αCD40 enabled a macrophage-independent T-cell response, engendering immune memory and durable complete remissions in a large fraction of mice harboring subcutaneous PDA tumors—and extending survival in KPC mice with spontaneous tumors. Both the use of nab-paclitaxel and the subcutaneous setting may be responsible for the differences in these observations, the subject of current investigations. Although of interest, we could not evaluate in Fc receptor-null mice whether Treg changes from αPD-1/αCTLA-4 were Fc receptor-dependent because the effects of agonistic αCD40 are lost in such mice (80).

In certain human cancers, tumor cell PD-L1 expression colocalizes with lymphocytic immune infiltrates, suggesting adaptive resistance by the tumor and the potential root of sensitivity to αPD-1 (33–35, 81). In the case of melanoma, tumor cell PD-L1 expression has been demonstrated to be dependent on CD8 A T cells and the secretion of IFNγ (36). In contrast, we found in our PDA mouse models harboring minimal intratumoral T cells, tumor cells express PD-L1 at moderate levels and tumor-associated DCs and macrophages express high levels of this inhibitory ligand. We further observed that PD-L1 expression in murine PDA is not dependent on T cells or IFNγ, indicating that PD-L1 tumor expression does not appear to be an adaptive response to immune pressure. These findings in the KPC model are corroborated by our observations in human PDA in which there was no spatial correlation between tumor PD-L1 expression and the presence of intratumoral CD8 A T cells. Tumor PD-L1 expression has been reported to be regulated by oncopogenes such as EGFR (82), suggesting that the regulation of PD-L1 in PDA may differ from the regulation of PD-L1 in other solid malignancies.

In summary, induction of T-cell immunity reduces resistance to PD-1 and CTLA-4 blockade and improves survival in pancreatic carcinoma. It is notable that four out of the five reagents that we used in these studies have an FDA-approved human homolog (gemcitabine, nab-paclitaxel, αPD-1, and αCTLA-4), which should facilitate clinical translation.

Disclosure of Potential Conflicts of Interest
E.J. Wherry has ownership interest (including patents) in Surface Oncology; is a consultant/advisory board member for Surface Oncology; and has provided expert testimony for Genentech/Roche. R.H. Vonderheide reports receiving commercial research grants from Roche and Pfizer. No potential conflicts of interest were disclosed by the other authors.

Authors’ Contributions
Conception and design: R. Winograd, E.J. Wherry, R.H. Vonderheide
Development of methodology: R. Winograd, K.T. Byrne, R.A. Evans, E.E. Furth, R.H. Vonderheide
Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): R. Winograd, K.T. Byrne, R.A. Evans, P.M. Odorizzi, A.R.I. Meyer, D.L. Bajor, C. Clendenin, E.E. Furth, R.H. Vonderheide
Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): R. Winograd, K.T. Byrne, R.A. Evans, P.M. Odorizzi, C. Clendenin, B.Z. Stanger, E.E. Furth, R.H. Vonderheide
Writing, review, and/or revision of the manuscript: R. Winograd, K.T. Byrne, R.A. Evans, E.E. Furth, E.J. Wherry, R.H. Vonderheide
Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): R. Winograd, A.R.I. Meyer, C. Clendenin, E.E. Furth, R.H. Vonderheide
Study supervision: R.H. Vonderheide

Acknowledgments
The authors thank Amy Ziober, Michael Feldman, Erin Delkava, Tim Chao, Andrew Rech, Mark Diamond, Steve Albleda, Jim Riley, Ellen Pure, and Anil Rustgi (Penn) and Dafna Bar-Sagi (New York University) for helpful discussions. Published OnlineFirst February 12, 2015; DOI: 10.1158/2326-6066.CIR-14-0215

www.aacrjournals.org Cancer Immunol Res; 3(4) April 2015 409
Grant Support
This study was supported by the NIH grants R01 CA169123 (to R.H. Vonderheide and B.Z. Stanger), T32 CA099140 (to R. Winograd), T32 HL07439 (to K.T. Byrnes), T32 HL007775 (to D.L. Bajoro), PS0 CA1061520 (to R.H. Vonderheide), U19 AI 082630, P01 AI 12521 (to E.J. Wherry), Stand Up To Cancer (to R.H. Vonderheide), Pancreatic Action Cancer Network-AACR (to R.H. Vonderheide), and Translational Center of Excellence in Pancreatic Cancer of the Abramson Cancer Center (to R.H. Vonderheide, C. Clendenin, and E.F. Farhi).

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received November 16, 2014; revised February 6, 2015; accepted February 6, 2015; published OnlineFirst February 24, 2015.

References
Extending Reach of Checkpoint Blockade in Pancreatic Cancer


Cancer Immunology Research

Induction of T-cell Immunity Overcomes Complete Resistance to PD-1 and CTLA-4 Blockade and Improves Survival in Pancreatic Carcinoma

Rafael Winograd, Katelyn T. Byrne, Rebecca A. Evans, et al.


Updated version
Access the most recent version of this article at:
doi:10.1158/2326-6066.CIR-14-0215

Supplementary Material
Access the most recent supplemental material at:
http://cancerimmunolres.aacrjournals.org/content/suppl/2015/02/12/2326-6066.CIR-14-0215.DC1

Cited articles
This article cites 82 articles, 38 of which you can access for free at:
http://cancerimmunolres.aacrjournals.org/content/3/4/399.full#ref-list-1

Citing articles
This article has been cited by 69 HighWire-hosted articles. Access the articles at:
http://cancerimmunolres.aacrjournals.org/content/3/4/399.full#related-urls

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, use this link
http://cancerimmunolres.aacrjournals.org/content/3/4/399.
Click on "Request Permissions" which will take you to the Copyright Clearance Center's (CCC) Rightslink site.