Efficacy of a Cancer Vaccine against ALK-Rearranged Lung Tumors

Claudia Voena1,2, Matteo Menotti1,2, Cristina Mastini1,2, Filomena Di Giacomo1,2, Dario Livio Longo1,3, Barbara Castella1,2, Maria Elena Boggio Merlo1,2, Chiara Ambrogio4, Qi Wang5, Valerio Giacomo Minero1,2, Teresa Poggio1,2, Cinzia Martinengo1,2, Lucia D’Amico1,2, Elena Panizza1,2, Luca Mologni6, Federica Cavallo1,7, Fiorella Altudra1,7, Mohit Butaney8,9, Marzia Capelletti8,9, Giorgio Inghirami1,2, Pasi A. Jänne8,9,10, and Roberto Chiarle1,2,5

Abstract

Non–small cell lung cancer (NSCLC) harboring chromosomal rearrangements of the anaplastic lymphoma kinase (ALK) gene is treated with ALK tyrosine kinase inhibitors (TKI), but the treatment is successful for only a limited amount of time; most patients experience a relapse due to the development of drug resistance. Here, we show that a vaccine against ALK induced a strong and specific immune response that both prophylactically and therapeutically impaired the growth of ALK-positive lung tumors in mouse models. The ALK vaccine was efficacious also in combination with ALK TKI treatment and significantly delayed tumor relapses after TKI suspension. We found that lung tumors containing ALK rearrangements induced an immunosuppressive microenvironment, regulating the expression of PD-L1 on the surface of lung tumor cells. High PD-L1 expression reduced ALK vaccine efficacy, which could be restored by administration of anti–PD-L1 immunotherapy. Thus, combinations of ALK vaccine with TKIs and immune checkpoint blockade therapies might represent a powerful strategy for the treatment of ALK-driven NSCLC. Cancer Immunol Res; 1–11. ©2015 AACR.

Introduction

Lung cancer is the leading cause of cancer-related mortality worldwide. In recent years, the identification of key genetic alterations in non–small cell lung cancer (NSCLC) has prompted the use of rationally targeted therapies, which showed unprecedented clinical benefits (1, 2). Approximately 5% to 6% of NSCLCs have chromosomal rearrangements of the anaplastic lymphoma kinase (ALK) gene that generate different chimeric proteins, such as EML4-ALK, TFG-ALK, and KIF5b-ALK (3–5). In all such fusions, constitutively active ALK acts as a potent tumor-igenic driver that activates downstream oncogenic signals, leading to increased cell proliferation and survival (4).

Experimental and clinical data show that ALK–rearranged NSCLCs respond to treatment with specific tyrosine kinase inhibitors (TKI), such as crizotinib (6, 7). Despite a high rate of initial response, the development of resistance to crizotinib almost invariably leads to tumor relapse and eventually to the patient’s death (8, 9). Next-generation ALK TKIs, such as ceritinib and alectinib, have been developed to overcome crizotinib resistance and can further extend survival in crizotinib-resistant patients (10–12). Resistance to ALK TKIs is mediated by point mutations of the ALK kinase domain, by ALK gene amplification, or by activation of other compensatory pathways, so-called bypass tracks, such as EGFR, c-KIT, c-MET, and IGF-R1 (8, 13–16). Thus, additional therapies to be combined with ALK TKIs are needed to further prolong remission or clinical response in NSCLC patients with ALK rearrangements.

Immunotherapy aimed at enhancing the immune response against tumor cells shows promising efficacy in a fraction of NSCLC (17, 18). In this context, the ALK protein has many features of an ideal tumor oncoantigen that can be exploited to design specific immunotherapies, such as a cancer vaccine. ALK is required for tumor survival and growth and expressed minimally in some nervous system cells (4). ALK is also antigenic in humans, as lymphoma patients with ALK rearrangements mount spontaneous B- and T-cell responses against the ALK protein, with measurable antibodies (20), cytotoxic T lymphocytes (CTL), and CD4+ T-helper effectors to ALK epitopes (21–24). A robust immune response to ALK is associated with a decreased risk of relapse in lymphoma patients (25). Our previous ALK vaccine in preclinical mouse models of lymphoma containing ALK rearrangements induced specific and potent immune responses that provided strong and durable tumor protection (19).
In the present study we test the efficacy of ALK vaccination in lung cancer. Grafted or primary mouse models of ALK-positive lung tumors showed that an ALK vaccine elicited a strong, ALK-specific CTL response in both mouse models, efficiently blocking tumor growth.

Materials and Methods

Cell lines and reagents

Human ALK-rearranged NSCLC cell lines H2228 (variant 3, E6; A20), DFCI032, and H3122 (variant 1, E13; A20) were obtained from the ATCC collection and were passaged for fewer than 6 months after receipt and resuscitation. These cell lines were further internally tested for the presence of EML-ALK rearrangement. The murine ASB-XIV cell line was purchased from Cell Lines Service (CLS) and was passaged for fewer than 6 months after receipt and resuscitation. The ALKTKI NVP-TAE684 was purchased from Axon Medchem and crizotinib (PF-02341066) was kindly gifted by P. Service (CLS) and was passaged for fewer than 6 months after receipt and resuscitation. These cell lines were further internally tested for the presence of EML-ALK rearrangement. The murine ASB-XIV cell line was purchased from Cell Lines Service (CLS) and was passaged for fewer than 6 months after receipt and resuscitation. These cell lines were further internally tested for the presence of EML-ALK rearrangement.

Mice

Strains of mice used in this study include K-RasL13G12V and Tg EGRF488R, as previously published (26, 27), and BALB/c mice (Charles River). Mice were handled and treated in accordance with European Community guidelines.

Generation of ALK transgenic mice

A cDNA fragment encoding EML4-ALK (variant 1, E13; A20) or TFG-ALK was ligated to the human SP-C promoter as well as to a polyadenylation signals (Supplementary Fig. S1A). The expression cassette was injected into pronuclear-stage embryos of FVB/N mice. The presence of the transgene was examined by PCR analysis with DNA from the tail of founder animals. Mice were handled and treated in accordance with European Community guidelines. Methods are further described in Supplementary Materials and Methods.

DNA vaccination and in vivo cytotoxicity assay

For DNA vaccination, 50 µg of pDEST or pDEST-ALK plasmids were used as previously described (19). The in vivo cytotoxicity assay was previously reported (19).

Antibody dosing for in vivo treatment

For CD4+ and CD8+ cell depletion, anti-CD4 (clone GK1.5) and anti-CD8 (clone 2.43) antibodies were purchased from BioXcell. For depletion, mice were injected i.p. with 100 µg of anti-CD4 or anti-CD8 at days −1, +5, +15, and +25.

For PD-1 blockade, anti–PD-1 (clone J43) and control anti-hamster polyclonal IgG for the in vivo experiments were purchased from BioXcell. Mice received 200 µg of each anti–PD-1 and anti–PD-L1 or 200 µg of anti-hamster IgG i.p. every 3 days for a total of 5 injections.

Magnetic resonance imaging

Magnetic resonance images (MRI) were acquired on a Bruker Avance 300 spectrometer operating at 7 T and equipped with a microimaging probe (Bruker Bio-Spin), as described in Supplementary Materials and Methods.

Histology and immunohistochemistry

For histologic evaluation, tissue samples were fixed in formalin, embedded in paraffin, stained, and visualized as previously described (19). T lymphocytes and regulatory T cells (Tregs) were quantified by measuring the number of CD3+ , CD8+ , CD4+ , and Foxp3+ cells, respectively, among the total tumor cells.

Intratumoral cell characterization

For flow cytometry analysis, lung cell infiltrate was obtained using the Lung Dissociation Kit (Miltenyi Biotec) following the manufacturer’s instructions. Cells were resuspended in phosphate buffer and stained with antibodies described in Supplementary Materials and Methods.

Statistical methods

Kaplan–Meier analyses for survival curves were performed with GraphPad Prism 5, and P values were determined with a log-rank Mantel–Cox test. Paired data were compared with the Student t test. P values of <0.05 were considered to be significant. Unless otherwise noted, data are presented as mean ± SEM.

Results

ALK vaccination elicits a specific cytotoxic response and prevents tumor growth in an orthotopic model of ALK-positive lung cancer

To test the efficacy of the ALK vaccine against lung cancer, we developed an orthotopic mouse model of ALK-positive lung cancer by ectopic expression of EML4-ALK in the syngeneic BALB/c murine lung cancer cell line ASB-XIV. We transduced ASB-XIV cells with a retroviral vector containing the EML4-ALK cDNA (variant 1) and GFP as a reporter. Protein expression in transduced ASB-XIV cells was comparable with EML4-ALK expression in human ALK-rearranged NSCLC cells (variant 1 in H3122 and 3 in H2228; Fig. 1A). ASB-XIV cells express MHC class I and thus are suitable for tumor immune studies (Fig. 1B). Within 3 weeks after i.v. injection of 5 × 105 ASB-XIV cells into the mouse tail vein, tumor nodules were detected in both lungs (Fig. 1E and F). We vaccinated BALB/c mice with a DNA plasmid coding for the intracytoplasmic domain of ALK (ref. 19; Fig. 1C).

ALK vaccine induced a strong ALK-specific immune response as measured by an in vivo cytotoxic assay (ref. 19; Fig. 1D). Ten days after the second vaccination, we injected EML4-ALK or GFP ASB-XIV cells. GFP ASB-XIV cells gave equal numbers of tumors in mice vaccinated with either a control or the ALK plasmid (Fig. 1E). In contrast, tumors of EML4-ALK ASB-XIV cells had impaired growth in ALK-vaccinated mice (Fig. 1F). Thus, ALK vaccination induced an ALK-specific immune response that efficiently prevented the growth of ALK-positive lung tumors.

ALK vaccination delays tumor growth and increases the overall survival of EML4-ALK–rearranged NSCLC Tg mice

To test the efficacy of the ALK vaccine as a therapy for primary lung tumors, we generated two transgenic (Tg) mouse models of ALK-driven lung cancers. Two rearrangements of ALK (EML4-ALK, variant 1, or TFG-ALK) were expressed under the human lung-specific surfactant protein-C (SP-C) promoter (Supplementary Fig. S1A), because human ALK-rearranged NSCLCs are often SP-C positive (28), and the expression of EML4-ALK by the SP-C promoter can induce efficient lung tumor formation in mice (29). Both transgenic mouse models expressed the ALK fusion selectively in lung epithelial cells, in amounts comparable with human NSCLC with rearranged ALK (Supplementary Fig. S1B–S1D), and rapidly developed multifocal...
ALK-positive tumors few weeks after birth, with 100% penetration (Supplementary Fig. S1E and S1F). Tumors were mainly well-differentiated adenocarcinoma growing as papillary, acinar, or more solid carcinoma (30). Ki-67 immunostaining showed that these tumors had a proliferation rate of 10.5%/64.1% for EML4-ALK and 8.5%/61.9% for TFG-ALK (Supplementary Fig. S1G). At 4 weeks of age, a few tumor nodules in both types of ALK mice (Supplementary Fig. S1H and S1I, left) were detected by MRI. Existing nodules rapidly expanded in volume and new nodules appeared in the lungs over time (Supplementary Fig. S1H and S1I, central and right plots). No tumor metastases were detected by examination of other organs with MRI or histology in ALK mice at any age, consistent with other constitutive or ALK-inducible mice (29, 31). Both EML4-ALK and TFG-ALK mice died within 50 weeks, with a mean survival of 33.25 weeks for EML4-ALK mice and 37 weeks for TFG-ALK mice (Supplementary Fig. S1L).

To test the efficacy of the ALK vaccine, we screened ALK mice by MRI to stratify them according to their tumor burden. ALK mice were vaccinated at 4 weeks of age, when tumors were detectable in the lungs (Supplementary Fig. S1H and S1I), according to the protocol in Fig. 2A. The ALK vaccine generated strong ALK-specific cytotoxic activity in both ALK models, comparable with wild-type (WT) littermates (Fig. 2B). In EML4-ALK mice, the average number of tumors detected in control mice was 58/66.0 by week 20, whereas ALK-vaccinated mice had only 16/63.5 at the same time point (Fig. 2C and D). Similar results were observed in TFG-ALK mice at 20 weeks of age (67/66.0 nodules in control mice compared with 20/63.5 nodules in vaccinated mice; Fig. 2E and Supplementary Fig. S2A). Correspondingly, the overall tumor burden calculated in terms of tumor volumes by serial MRI was significantly lower in ALK-vaccinated mice than in control mice (Supplementary Fig. S2B and S2C). Survival of ALK-vaccinated mice was
significantly extended by at least 18 weeks in EML4-ALK and by 12 weeks in TFG-ALK mice (Fig. 2F and G). The ALK vaccine was still efficacious against larger tumors in older mice (Supplementary Fig. S2D). Thus, ALK-DNA vaccination was a potent controller of growth of primary ALK-rearranged lung tumors.

ALK-DNA vaccination increases the number of intratumoral T cells and requires CD8+ T lymphocytes

Next, we examined how the ALK vaccine shaped the intratumoral immune infiltrate. The ALK vaccine increased the number of intratumoral T cells in both EML4-ALK and TFG-ALK mice, which was associated with a reduced tumor size (Fig. 3A and B). Both CD4+ and CD8+ T cells were significantly increased in ALK-vaccinated mice (Fig. 3C). In ALK-vaccinated mice, tumor-infiltrating T cells had a significantly higher CD8+ :CD4+ ratio compared with controls, due to the DNA vaccine preferentially stimulating a CD8+ T-cell immune response (Fig. 3C; ref. 32). We also observed an increase in intratumoral Tregs (Fig. 3D and E), suggesting that the ALK vaccine induces both T eff cells and Tregs, as described for other tumor vaccines (33). Nonetheless, the ratio of CD8+ :Foxp3+ was higher in vaccinated mice than in control mice (Fig. 3E).

To confirm that the ALK vaccine required T eff for its antitumor functions, we used repeated injections of antibodies specific for CD4+ and CD8+ T cells to deplete them in the orthotopic model based on EML4-ALK ASB-XIV cells (Fig. 3F). The depletion of CD8+ T cells, but not CD4+ T cells, significantly reduced the ALK vaccine efficacy (Fig. 3G and H). Therefore, in mice bearing ALK-positive tumors, ALK vaccination elicited a cytotoxic response largely mediated by CD8+ T cells. However, in mice depleted of CD8+ T cells, the vaccine still appeared to retain some activity, suggesting that other factors may be involved in the immune response elicited by the vaccine.

Immunosuppressive lung microenvironment in ALK-rearranged lung cancer

We showed that the ALK vaccine stimulates a potent immune response against ALK-rearranged lung tumors. However, the
ALK vaccine did not cure the mice, which died after a delay in tumor growth (Fig. 2). Because ALK was still expressed in late tumors, we asked whether the efficacy of the ALK vaccine would diminish over time due to an immunosuppressive microenvironment that progressively develops in lung tumors with ALK rearrangements. Indeed, oncogenic activation of EGFR in lung cancers induces an immunosuppressive lung microenvironment by induction of PD-L1 expression on the surface of tumor epithelial cells (34).

First, we better characterized the immune infiltrate in mice bearing ALK lung tumors and observed that overall the percentage of B and T lymphocytes, natural killer (NK) cells, and granulocytes was comparable between WT and EML4-ALK mice (Supplementary Fig. S3A–S3D). However, T cells in tumor-bearing EML4-ALK mice displayed a significantly higher expression of PD-1 on both CD4⁺ and CD8⁺ T cells (Fig. 4A and Supplementary Fig. S3E), and PD-1⁺CD3⁺ T cells also expressed other T-cell–inhibitory molecules such as LAG-3 and TIM-3 in higher amounts (Supplementary Fig. S3F). These findings were further extended by interrogating gene-expression profiling data from larger series of human NSCLC with different oncogenic mutations. By gene set enrichment analysis, we found lower expression of tumor-infiltrating T-cell–related molecules in EML4-ALK NSCLC compared with EGFR-mutated, K-RAS–mutated, or ALK/RAS/EGFR–nonmutated NSCLC (Fig. 4C). In particular, in ALK-rearranged tumors, we...
observed significant depletion of T-cell receptor (TCR)-related molecules, such as TCRβ chain, CD3ε, CD3γ, CD3ζ, and Lck, of the T-cell costimulatory molecules ICOS and CD28, as well as of CD80 and CTLA-4 (Supplementary Fig. S4A–S4D).

Blockade of the PD-1/PD-L1 pathway restores ALK vaccine efficacy against tumor cells with high levels of PD-L1

We asked whether oncogenic ALK could also regulate PD-L1 expression on lung tumors. Tumors derived from EML4-ALK mice had higher levels of PD-L1 expression than tumors originating in mice carrying other NSCLC recurrent mutations, such as the K-RasG12V (26) and EGFRL858R (27) mice (Supplementary Fig. S5A). Next, we analyzed PD-L1 expression by flow cytometry and showed that tumor epithelial cells (CD45- /EpCAM+) and associated hematopoietic cells (CD45+) in EML4-ALK mice expressed PD-L1 (Supplementary Fig. S5B). To determine whether ALK oncogenic activity directly controlled PD-L1 expression in NSCLC, we treated three ALK-rearranged NSCLC cell lines (H3122, H2228, and DFCI032) with crizotinib to inhibit ALK activity (Fig. 5A and Supplementary Fig. S5C). Expression of PD-L1 was detectable in all ALK-rearranged cell lines tested and was associated with oncogenic activity directly controlled PD-L1 expression in all three cell lines (Fig. 5B and Supplementary Fig. S5D). Consistently, PD-L1 mRNA was also downregulated (Fig. 5C and Supplementary Fig. S5E). To further confirm that PD-L1 expression was driven by ALK activity, and to exclude the possibility that PD-L1 downregulation was mediated by crizotinib inhibition of MET, ROS1, or other off-targets, we knocked down EML4-ALK by a doxycycline-inducible shRNA construct (Fig. 5D). Vaccinated mice were inoculated with control EML4-ALK ASB-XIV cells or EML4-ALK ASB-XIV cells expressing high PD-L1, and we observed similar intensity of expression by flow cytometry in EML4-ALK ASB-XIV (Fig. 5D). We reasoned that ALK vaccine efficacy could be hampered when target tumor cells express more PD-L1. We engineered EML4-ALK ASB-XIV cells to express more PD-L1 than parental cells by transduction with a lentivirus containing a murine PD-L1 construct (Fig. 5D). Vaccinated mice were injected with control EML4-ALK ASB-XIV cells or EML4-ALK ASB-XIV cells expressing high PD-L1. In the presence of high PD-L1 expression, the ALK vaccine was less effective in preventing lung tumor growth (Fig. 5E), suggesting that the function of ALK-specific T eff cells was modulated by the amount of PD-L1 on the surface of target lung tumor cells.
To test whether administration of antibody to PD-1 could restore a full efficacy of the ALK vaccine, we treated mice with anti–PD-1 or control IgG (Supplementary Fig. S6A). The treatment with anti–PD-1 alone, as well as control IgG, did not have significant effect on tumor growth and mice developed tumors comparable with controls. In contrast, anti–PD-1 treatment almost completely restored the efficacy of the ALK vaccine (Fig. 5F). These results are consistent with findings that immune checkpoint therapy restores an adaptive immune response best in patients with high PD-L1 expression (37, 38).

To show that blockade of the PD-L1/PD-1 immune checkpoint was effective with physiologic expression of PD-L1, we tested anti–PD-1 treatment in EML4-ALK mice (Supplementary Fig. S6B). We observed a stabilization of tumors immediately at the end of treatment (Fig. 5G) followed by a slower growth rate as compared with control mice (Fig. 5H). These data suggest that immune checkpoint blockade therapy could be efficacious in the physiologic tumor microenvironment of ALK-rearranged lung tumors.

ALK vaccination is effective in combination with ALK TKIs

Crizotinib treatment of NSCLC patients with ALK rearrangements has had success in clinical trials, supporting the use of ALK TKIs as main therapy for NSCLC (39). A combination of ALK TKIs with the ALK vaccine could be an attractive therapeutic possibility for NSCLC patients. In this context, ALK TKIs could reduce the tumor burden to facilitate the activity of the ALK vaccine. We tested this combination in our ALK mouse model...
ALK mice were treated with crizotinib (PF-02341066) for 2 weeks (100 mg/kg) and concurrently vaccinated with the ALK or control vaccine (Fig. 6A). The immune response elicited by the vaccine was not affected by administration of crizotinib, as an equally strong ALK-specific cytotoxic immune response in vivo was also detected in ALK-vaccinated mice treated with crizotinib (Fig. 6B). As expected, treatment with crizotinib induced the regression of tumors in both groups within 2 weeks (Fig. 6C, left and central plots; 6D). At 6 weeks from treatment suspension, tumors relapsed at the same sites in crizotinib-treated mice (Fig. 6C, top right plots), whereas the combination of crizotinib and ALK vaccine delayed tumor recurrence (Fig. 6C, bottom right plots). Indeed, mice treated with crizotinib showed relapses and new tumors 10 weeks after treatment suspension, whereas in ALK-vaccinated mice, relapses and new tumors were less numerous and significantly smaller (Fig. 6D and E). Similar results were obtained when EML4-ALK mice were vaccinated during treatment with TAE684 (25 mg/kg; Supplementary Fig. S7A–S7C). Thus, the ALK vaccine might be efficiently combined with ALK TKI treatment to delay tumor relapse after crizotinib suspension.

**ALK vaccination prevents growth of tumors expressing crizotinib-resistant ALK mutations**

Human NSCLC patients treated with ALK TKI almost invariably develop resistance. L1196M, C1156Y, and F1174L are common ALK mutations found in patients relapsing under treatment with crizotinib (8, 12, 13). Point mutations in the ALK kinase...
domain have the potential to alter the antigenicity of ALK as they can modify its protein structure. To test the activity of the ALK vaccine against these mutants, we transduced ASB-XIV cells with a retroviral vector containing either the EML4-ALK WT or the EML4-ALK mutants (Fig. 7A). Control mice injected with ASB-XIV cells expressing the L1196M, C1156Y, or F1174L EML4-ALK mutants rapidly developed tumors in the lungs, whereas ALK vaccination almost completely prevented tumor growth of EML4-ALK WT and all mutants (Fig. 7B–E). Therefore, the ALK vaccine is not only efficacious against the EML4-ALK WT but also against some of the most common EML4-ALK mutants that develop in patients during treatment with crizotinib or ceritinib.

Discussion

In this work, we extended our previous findings on the efficacy of a DNA ALK vaccine against ALK-positive lymphoma to ALK-rearranged lung cancers. Compared with our previous work, we showed that the ALK vaccine is active not only in tumor grafts but also in primary ALK-rearranged NSCLC. Because the SP-C promoter is active since embryonic development (40), these mice are likely tolerant to human ALK. Thus, an important advance from this work is the demonstration that an ALK vaccine can overcome tolerance in mice.

In addition, we showed that the ALK vaccine could be combined with either ALK TKI treatment or the anti–PD-1 antibody. These combinatorial therapies make the ALK vaccine attractive for potential clinical use. Current therapies for ALK-rearranged NSCLC, based on crizotinib or next-generation ALK TKIs, achieve a clinical response by arresting tumor progression or inducing tumor regression. However, all patients eventually relapse and die due to development of TKI resistance (11, 41).

In this work, we set the stage for the application of an ALK vaccine to further extend progression-free survival in NSCLC patients. The ALK vaccine induced a strong systemic and intratumoral immune response in mouse models of ALK-rearranged NSCLC, significantly reducing tumor growth and extending survival of treated mice, regardless of the type of ALK translocation (EML4-ALK or TFG-ALK). Simultaneous treatment during vaccination with crizotinib or TAE684 did not affect the ALK immune response achieved by the vaccine. Thus, these data indicate the feasibility of administering an ALK vaccine to NSCLC patients during ALK-TKI treatment, possibly when the response is maximal in terms of tumor burden reduction.
Additional advantages of such a combination could stem from the potential activity of ALK TKIs in the regulation of the tumor immune microenvironment. We showed that the oncogenic activity of ALK directly regulated PD-L1 expression of the surface of tumor cells. High PD-L1 expression impaired the immune response against ALK elicited by the vaccine (Fig. 5). Therefore, PD-L1 downregulation by ALK TKI treatment could relieve the inhibitory feedback on intratumoral T cells and facilitate ALK-specific immune responses. Tumor cell death induced by ALK TKIs could release additional tumor neoantigens, including ALK itself, and thus enhance antitumor immune response (42, 43). Further investigation to elucidate the effect of ALK TKIs on the tumor microenvironment is required, but it is intriguing that studies in mouse models and metastatic melanoma patients showed an enhanced antitumor immune response after treatment with the selective B-RAF inhibitor ( vemurafenib) alone, or in combination with MEK inhibitors (44, 45).

The immune microenvironment in ALK-rearranged tumors could therefore be a critical factor in the ALK-specific immune responses. We presented data indicating that ALK-rearranged mice indeed progressively develop an immunosuppressive tumor microenvironment similar to that induced in mice by oncogenic EGFR (34). Compared with WT mice, ALK-rearranged lungs accumulated higher numbers of PD-1+ T cells that also expressed the exhaustion markers TIM-3 and LAG-3, and showed increased numbers of tumor-infiltrating Tregs. ALK-rearranged NSCLC patients also showed a likely immunosuppressive microenvironment in the lungs with reduced tumor-infiltrating T cells.

In ALK-vaccinated lungs, the tumor-infiltrating Tregs were increased, and we detected a population of intratumoral CD8+ T cells with high expression of PD-1 (Supplementary Fig. S8A and S8B), which we interpreted as exhausted CD8+ T cells that had been elicited by the ALK vaccine to recognize the ALK antigen (46). In mice with advanced tumors, the ALK vaccine elicited a weaker ALK-specific cytotoxic response (Supplementary Fig. S8C) and decreased antitumor activity (Supplementary Fig. S2D). In these settings, Treg depletion by an antibody to CD25 could partially restore the impaired cytotoxic activity generated by the ALK vaccine (Supplementary Fig. S8C), indicating that Tregs could also play a critical role in the immunosuppressive tumor environment seen in ALK-rearranged lung tumors. Similarly, the restoration of the ALK vaccine efficacy by administration of antibody to PD-1 in high PD-L1 EML4-ALK ASB-XIV xenografts (Fig. 5) suggests that blockade of immune checkpoint molecules could powerfully potentiate the ALK vaccine.

Overall, these data suggest that combination therapy of ALK TKIs and ALK vaccine could work efficiently in the clinical setting to generate a strong and long-lasting immune response to ALK in NSCLC. The benefit from combined ALK TKI and ALK vaccine therapy can be enhanced by additional immunotherapies, such as anti–PD-1/PD-L1 and anti–CTLA, to block immune checkpoints (17, 18) or through Treg depletion by antibodies to CD25 (47). Thus, the development of an ALK vaccine for clinical use, together with additional immunotherapeutic tools, provides exciting therapeutic options for the treatment of ALK-rearranged NSCLC.

Disclosure of Potential Conflicts of Interest
P.A. Janne is a consultant/advisory board member for Chugai, Pfizer, and Roche. No potential conflicts of interest were disclosed by the other authors.

Authors’ Contributions
Conception and design: C. Voena, G. Inghirami, R. Chiarle
Development of methodology: C. Voena, C. Mastini, F. Di Giacomo, C. Ambrogio, L. D’Amico, E. Panizza, R. Chiarle
Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): C. Voena, M. Menotti, C. Mastini, F. Di Giacomo, D.L. Longo, R. Castella, M.E. BoggioMerlo, C. Ambrogio, V.G. Minero, T. Poggio, C. Martinengo, F. Cavallo, F. Altunba, M. Butaney, M. Capelletti, P.A. Janne, R. Chiarle
Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): C. Voena, M. Menotti, C. Mastini, F. Di Giacomo, R. Castella, M.E. BoggioMerlo, C. Ambrogio, Q. Wang, V.G. Minero, C. Martinengo, M. Butaney, R. Chiarle
Writing, review, and/or revision of the manuscript: C. Voena, D.L. Longo, P.A. Janne, R. Chiarle
Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): C. Voena, F. Cavallo, M. Butaney
Study supervision: R. Chiarle
Other (provided reagents): L. Mologni

Acknowledgments
The authors thank Glenn Dranoff for helpful discussions, Maria Stella Scalzo for technical support, Flavio Cristofani for housing and care of mice, Carlo Gambacorti-Passerini for kindly providing critical reagents, Silvio Aime for the Molecular Imaging Facility, and Sharmila Fagoonee and Maddalena Iannicella for technical help in generation of transgenic mice.

Grant Support
This work has been supported by grants FP7 ERC-2009-StG (proposal no. 242965—"Lunely") to R. Chiarle; Associazione Italiana per la Ricerca sul Cancro (AIRC) grant IG-12023 to R. Chiarle; Koch Institute/DFCC Bridge Project Fund to R. Chiarle; Ellison Foundation Boston to R. Chiarle; the Grant for Oncology Innovation by Merck-Serono to R. Chiarle; Worldwide Cancer Research (former AICR) grant 12-0216 to R. Chiarle; and R01 CA136851 to P.A. Janne.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received April 2, 2015; revised June 2, 2015; accepted July 23, 2015. Published OnlineFirst September 29, 2015.
Cancer Immunology Research

Efficacy of a Cancer Vaccine against ALK-Rearranged Lung Tumors

Claudia Voena, Matteo Menotti, Cristina Mastini, et al.

Cancer Immunol Res  Published OnlineFirst September 29, 2015.

Updated version
Access the most recent version of this article at:
doi:10.1158/2326-6066.CIR-15-0089

Supplementary Material
Access the most recent supplemental material at:
http://cancerimmunolres.aacrjournals.org/content/suppl/2015/10/17/2326-6066.CIR-15-0089.DC1

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.